PRO-LAB DIAGNOSTICS

MICROBANK®

World Wide Performance Portfolio

With 2D Barcodes

5th Edition





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Microbank® Introduction

Microbank® with 2D DataMatrix GS1-compliant barcode

Microbank®, a proven system for the long-term storage and retrieval of bacterial and fungal cultures has details of successful referenced performance data collected over 28 years from reference centres and laboratories around the world detailing data ranging from 2 - 15 years. Millions of vials have been used worldwide and this continues.

In addition to its now infamous presentation, Microbank® has been enjoying success in North America with a new 2D DataMatrix GS1-compliant barcoded version. Now established for some time, we are pleased to advise the arrival of the new feature in Europe. The concept is simple: the unique barcode is incorporated into the vial label area, leaving ample space for additional labelling, and on the storage box.

To enhance your experience with the 2D barcodes, we are also pleased to offer the following: your own Penguin Barcode Reader, Data Storage Device, and a personalised Microbank® Assets Web Database providing the full package of everything you need to get started and to ensure easy location of any stored culture with instant access to its source history. You can bet there will be another “Mug” in the series out very soon as well to add to your collection.

Microbank® has been manufactured exclusively by Pro-Lab Diagnostics for over 28 years. Only Microbank® offers up to 28 years of successful storage; data is available in the Microbank® Worldwide Performance Portfolio making it the choice for laboratories worldwide and beyond. Yes, beyond: we have a bacterial collection in outer space developed specifically for a rather well-known organisation!

The long-term storage of microorganisms is a significant challenge in microbiology. Microbank® with unique 2D barcodes offers a platform that utilises porous glass beads and a specially formulated cryopreservative for storage at low temperatures. The additional feature of a unique 2D barcode on each box and each vial facilitates easier documentation and retrieval of your isolates. This format coupled with the Microbank® Cryoblock offers the least possibility of disturbance to your organism, yet permits ready and rapid access.

Each Microbank® vial contains approximately 25 sterile coloured beads (single colour) and the specially formulated cryopreservative unique to Microbank®, allowing storage from -20°C to -70°C. The specially treated beads are of a porous nature allowing microorganisms to readily adhere onto the bead surface. After inoculation the Microbank® vials are kept frozen for extended storage. When a fresh culture is required, a single bead is easily removed from the Microbank® vial and used to directly inoculate a suitable culture medium.

The unique 2D barcodes on the box and vials provide you with a freezer-safe, durable label that can be scanned into your Laboratory Information System or Assets Web Database with ease. This feature not only removes the risk of transcription errors but has the added advantage of making retrieval from the freezer a simple task.

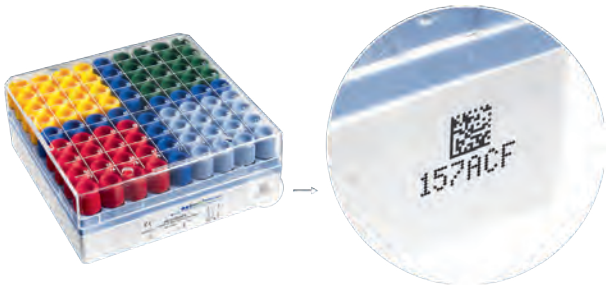
2D Explained

Pairing the unique 2D DataMatrix barcode on the storage box to the vials supplied will allow for better traceability of cultures as you build your database. 2D barcodes use patterns of squares, hexagons, dots, and other shapes to encode data. Because of this structure, 2D barcodes can hold data up to 2000 characters, and are very small. The data is encoded based on both the vertical and horizontal arrangement of the pattern, and is read in two dimensions.



Linear or 1D barcodes, like the UPC code commonly found on consumer goods, use a series of variable-width lines and spaces to encode. Whilst acceptable in many applications they hold just a few dozen characters, and generally get physically longer as more data is added; use can be typically limited to 8-15 characters, causing problems on small labels where additional data may also be required.

2D barcodes have increasingly been used in supply chain and manufacturing applications with medical equipment and diagnostic industries, where companies have been tasked with providing a large amount of product tracking information on some very small items. For example, the U.S. FDA's UDI rules require several pieces of manufacturing information to be included on certain types of medical devices. That data could be easily encoded on very small 2D barcodes.





"The type of barcode selected requires careful consideration and is dependent on quality, ease of use, and the specific requirements of the application, including the type and amount of data that is required to be, the size of the asset/item, and how and where the code will be scanned."

"The adoption of the 2D DataMatrix GS1-Compliant Barcode on the Microbank® vial offers an efficient system available."

Microbank® Key Features



- Proven performance over 28 years with a collection of published storage data papers internationally.
- References available for storage at -20°C to -70°C.
- Unique cryopreservation solution suitable for aerobic, anaerobic, fastidious bacteria, and fungal cultures.
- Unique treated beads to enhance bacterial adhesion
- Available in five standard colours.
- Standard pack of 80 supplied in durable storage box
- Custom pack sizes available for large collections.
- Multiple colour coding option available (120 available).
- Industry-standard cryovials with large writing area and triple-depth external threaded tube which reduces the possibility of contamination and improves handling.

Large 2 ml vials

To offer improved handling, inoculation, and retrieval.

Available in five colours

To provide laboratories with a system to colour code different bacterial and fungal species.

2D DataMatrix GS1-compliant Barcode

Larger writing area

Allows for complete coding and reference data.

Chemically treated beads

Improved bacterial adhesion.

Industry-standard

Robust cryovial can withstand snap freezing with liquid nitrogen.

Specially formulated preservative

Ensures longer survival of fastidious bacteria and higher quantitative recoveries.

Product Availability

Microbank®

Advanced presentation of 80 vials supplied in a plastic freezer box manufactured from durable plastic with "see through" lids, number locator printed screens, and tube collection device.

MICROBANK® WITH 2D DATAMATRIX GS1-COMPLIANT BARCODE

PRODUCTS	CODE	SIZE
Microbank® (with 2D barcode) - Blue	PL.170C/B	80 Vials
Microbank® (with 2D barcode) - Green	PL.170C/G	80 Vials
Microbank® (with 2D barcode) - Red	PL.170C/R	80 Vials
Microbank® (with 2D barcode) - Yellow	PL.170C/Y	80 Vials
Microbank® (with 2D barcode) - Light Blue	PL.170C/LB	80 Vials
Microbank® (with 2D barcode) - Mixed (16 x each colour)	PL.170C/M	80 Vials

MICROBANK® SPECIAL PRESERVATION ONLY

Supplied in the same format as traditional Microbank® but with specially formulated broth only.

PRODUCTS	CODE	SIZE
Microbank® - Blue	PL.173/B	80 Vials
Microbank® - Green	PL.173/G	80 Vials
Microbank® - Red	PL.173/R	80 Vials
Microbank® - Yellow	PL.173/Y	80 Vials
Microbank® - Light Blue	PL.173/LB	80 Vials
Microbank® - Mixed (16 x each colour)	PL.173/M	80 Vials

MICROBANK® DRY

Supplied in the same format as traditional Microbank® but without the specially formulated cryopreservation solution.

PRODUCTS	CODE	SIZE
Microbank® - Blue	PL.172/B	80 Vials
Microbank® - Green	PL.172/G	80 Vials
Microbank® - Red	PL.172/R	80 Vials
Microbank® - Yellow	PL.172/Y	80 Vials
Microbank® - Light Blue	PL.172/LB	80 Vials
Microbank® - Mixed (16 x each colour)	PL.172/M	80 Vials

MICROBANK® ACCESSORIES

Freezer storage boxes are available which are suitable for collection and organisation in low-temperature freezers. Available in blue and red, each freezer box will hold 81 Microbank® vials. Also available is the Cryoblock for maintaining low temperatures while working with Microbank® vials on the laboratory bench.

PRODUCTS	CODE	SIZE
Freezer Storage Box - Blue	PL.169/B-1	Each
Freezer Storage Box - Red	PL.169/R-1	Each
Freezer Storage Box - Blue	PL.169/B-4	4 Pack
Freezer Storage Box - Red	PL.169/R-4	4 Pack
Cryoblock	PL.155-1	20 Well
Insulated Base & Lid	PL.156	2 Pack
Laminated Log Book	PL.165	12 Charts
Aluminium Cryocanes	PL.166	12 Canes
Microbank® - 2D Barcode Reader	PL.168	Each

Finally, our product mascot simply can't go without mention.

This cute little guy has become synonymous with the Microbank® system; he appears on mugs (9 in the series now!), key rings, socks, bottle openers, USB sticks, Christmas trees and he even has his own calendar monitoring his travels. If you have any pictures for inclusion in the calendar, or if you need a new penguin to take on your travels please email:

uksupport@pro-lab.co.uk





Microbank® Instructions For Use

Intended Use

Microbank® is a ready-to-use system designed for the long-term storage and retrieval of bacterial and fungal isolates.

Summary and Explanation

The long-term storage of microorganisms is a significant challenge in microbiology. Microbank® with unique 2D barcodes offers a platform that utilises porous glass beads and a specially formulated cryopreservative for storage at low temperatures. The additional feature of a unique 2D barcode on each box and each vial facilitates easier documentation and retrieval of your isolates. This format coupled with the Microbank® Cryoblock offers the least possibility of disturbance to your organism, yet, permits ready and rapid access.

*'The unique 2D barcodes on the box and vials provide you with a freezer-safe, non-erasable label that can be **scanned into your Laboratory** Information System with ease.'*

Description

Each Microbank® vial contains approximately 25 sterile coloured beads (single colour) and the specially formulated cryopreservative. The specially treated beads are of a porous nature allowing microorganisms to readily adhere onto the bead surface. After inoculation the Microbank® vials are kept at -70°C for extended storage. When a fresh culture is required, a single bead is easily removed from the Microbank® vial and used to directly inoculate a suitable culture medium.

The unique 2D barcodes on the box and vials provide you with a freezer-safe, non-erasable label that can be scanned into your Laboratory Information System with ease. This feature not only removes the risk of transcription errors but has the added advantage of making retrieval from the freezer a simple task.

Procedure

Scan the 2D barcode on the Microbank® box and record any relevant information in your freezer storage programme.

Scan the 2D barcode on the side of the Microbank® vial and record relevant information for each organism to be stored.



Inoculate the cryopreservative fluid with young colonial growth (18-24 hours) picked from a pure culture to approximately a 3-4 McFarland standard.



Close the Microbank® vial tightly and invert 4-5 times. Do not vortex.

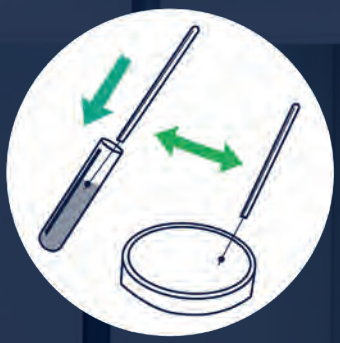


At this point the microorganisms will be bound to the porous beads. The excess cryopreservative should be well aspirated leaving the inoculated beads as free of liquid as possible.



Using your freezer programme find the location of the isolate you wish to work with. Scan the 2D barcode on the Microbank® vial to ensure the correct isolate is being retrieved.

Under aseptic conditions, open the cryovial and using a sterile needle or forceps remove one coloured bead.



Close the vial finger tight and return as soon as possible to low-temperature storage. Excessive changes in temperature reduce the viability of the organisms.

The inoculated bead may then be used to directly streak on to solid medium or may be dropped into an appropriate liquid medium.

*When used as recommended, each cryovial will store approximately **25** identical potential cultures.*

Limitations

Microbank® is offered solely as a means of providing long-term storage of bacterial and fungal isolates. Aseptic technique should be practised at all times to ensure continued integrity of the stored isolate.

Beads should never be returned to the Microbank® vial for any reason.
Microbank® vials are supplied in a variety of colours. These colours do not imply any change in the product's function. The different colours are provided so that the user can utilize them for their own tracking purposes.

- *Microbank® should not be used if any of the following conditions are present before inoculation.*
- *The vial shows any evidence of leakage (loss of cryopreservative).*
- *There is excess turbidity in the cryopreservative suggesting contamination.*
- *The expiration date on the outer label has elapsed.*

Safety Precautions

Observe biohazard precautions when preparing new or discarding used Microbank® vials. When storing Microbank® vials in liquid nitrogen the following precautions should be taken:

- *Always use the appropriate safety equipment.*
- *Microbank® vials should only be placed in the vapour phase of the liquid nitrogen.*
- *Ensure that the threads of the Microbank® vial and screw cap are completely dry before closing.*
- *Ensure the Microbank® vial cap is tight. Do not over tighten.*

Presentation

Microbank® is packaged in shelf packs of 80 vials.

Storage

Before use, unused Microbank® vials may be stored at room temperature. Stored under these conditions Microbank® vials may be used up to the date of expiration shown on the product label.

References

White DJ. and Sand, R.L. 1985. Medical Laboratory Sciences 42:289-290(UK).
Feltham et al. 1978. Journal of Applied Bacteriology. 44:313-316.
Nagel, J.G. and Cunz, L.J. 1971. Applied Microbiology, 23(4):837-838

Please refer to IFU PL.170C at www.pro-lab.com or www.pro-lab.co.uk for full details

Anaerobe Reference Unit Microbank® Storage Trial 5 Years

Dr J. Brazier, Dr V. Hall.
Anaerobic Reference Centre, Cardiff, UK.

Methods and Materials

One bead from each vial will be inoculated onto Fastidious Anaerobe agar (Lab M, Bury) with 5% horse blood, and spread for single colonies. Plates will be incubated promptly in an anaerobic chamber (Concept 300 or Concept Plus) at 37°C for 48 hours. On removal from the chamber, cultures will be examined for growth consistent with the intended isolate. Cultures yielding no growth will be incubated for a further 3 days and re-examined.

Results

Growth will be recorded as + (intended isolate recovered) or - (isolate not recovered). Original inocula were not standardised, therefore, quantitation of growth would be fairly meaningless. Besides, for our purposes, density of growth is unimportant as long as the isolate is recoverable.

ORGANISM	GROWTH	ORGANISM	GROWTH
A. gerencseriae	+	Actinomyces sp.	+
A. israelii	+	Actinomyces sp.	+
A. israelii	+	Actinomyces viscosis	+
A. naeslundii	+	B. fragilis	+
A. odontolyticus	+	B. ovatus	+
A. turicensis	+	B. splanchnicus	+
A. naeslundii	+	B. splanchnicus	+

ORGANISM	GROWTH	ORGANISM	GROWTH
B. thetaiotaomicron	+	C.sordellii	+
B. vulgaris	+	C.sporogenes	+
B.distasonis	+	C.tetani	+
B.distasonis	+	Camp .ureolyticus	+
B.fragilis	+	Camp. ureolylicus	+
B.fragilis	+	Camp. gracillis	+
A. naeslundii	+	Campylobacter recta	+
B.ovatus	+	B. vulgaris	+
B.thetaiotaomicron	+	Campylobacter spp.	+
B.uniformis	+	Clostridium perfringens	+
Bacteroides thetaiotaomicron	+	E. aerofaciens	+
Bif. Animalis group	+	E. lentum	+
Bif. Longum	+	E. lentum	+
Bifodobacterium spp.	+	Eu. lentum	+
Bifodobacterium spp.	+	Eubacterium aerofaciens	+
Bilophilia wadsworthia	+	F. naviforme	+
Bilophilia wadsworthia	+	F. necrophorum	+
C.bifermentans	+	F. necrophorum	+
C.butricum/beijerinckii	+	F. nucleatum	+
C.cadaveris	+	F. nucleatum	+
C.cadaveris	+	F. nucleatum	+
C.clostridioforme	+	F. russii	+
C.difficile	+	F. varium	+
C.novyi type A	+	Fusobacterium necrophorum	+
C.paraputrificum	+	L. acidophius	+
C.perfringens	+	L. acidophius	+
C.ramosum	+	Lactobacillus acidophilus	+
C.septicum	+	Peptostrep .asaccharolyticus	+

ORGANISM	GROWTH	ORGANISM	GROWTH
Peptostrep. Asaccharolyticus	+	Streptococcus mutans	+
Peptostrept. anaerobius	+	Veillonelle parvula	+
Peptostrept. Magnus	+	Veillonelle parvula	+
Peptostrept. micros	+		
Peptostrept. micros	+		
Peptostrept.anaerobius	+		
Peptostreptococcus productus	+		
Peptostreptococcus magnus	+		
Peptostreptococcus productus	+		
Peptostreptococcus sp.	+		
Peptostreptococcus sp.	+		
Peptostreptococcus sp.	+		
Porph. endodontalis	+		
Porph. levii	+		
Porphyromonas asaccharolytica	+		
Porphyromonas sp.	+		
Prev. denticola	+		
Prev. melaninogenica	+		
Prev. melaninogenica productus	+		
Prev. oris	+		
Prevotella loescheii	+		
Prevotella spp.	+		
Prevotella spp.	+		
Prop. acnes	+		
Prop. granulosum	+		
Prop. propionicum	+		
Propionibacterium acnes	+		
Staphylococcus saccharolyticus	+		

Anaerobe Reference Unit Microbank® Storage Trial 7 Years

Dr J. Brazier, Dr V. Hall.
Anaerobic Reference Centre, Cardiff UK.

Methods and Materials

Microbank® vials were inoculated with clinical isolates of obligately anaerobic bacteria referred to the Anaerobe Reference Unit for confirmation of identity and were frozen at -80°C for seven years. Isolates for study (n=100) were selected to represent the range of species commonly isolated from clinical material. One bead from each vial had been removed for culture after five years. Beads may have been removed on other occasions.

One bead from each vial was placed aseptically onto Fastidious Anaerobe agar (IDG, Bury) containing 5% horse blood. Plates were spread for single colonies and promptly incubated in an anaerobic chamber (Concept Plus) for 48 hours.

On removal from the chamber, plates were examined for growth of colonies characteristic of the intended isolates. If growth was not apparent at this stage, plates would be re-incubated for a further three days but, in the event, this was not necessary.

Results

All 100 isolates were cultured after anaerobic incubation for 48 hours. Isolates examined were:

Actinomyces sp. (2)	(2)
Actinomyces gerencseriae	
Actinomyces israelii	(2)
Actinomyces naeslundii	(2)
Actinomyces odontolyticus	
Actinomyces turicensis	
Actinomyces viscosis	
Bacteroides distasonis	(2)
Bacteroides fragilis	(4)
Bacteroides ovatus	(2)
Bacteroides splanchnicus	(2)
Bacteroides thetaiotaomicron	
Bacteroides uniformis	

Bacteroides vulgaris	
Bifidobacterium spp.	(2)
Bifidobacterium animalis group	
Bifidobacterium longum	
Clostridium bifermentans	
Clostridium butyricum / beijerinckii	
Clostridium cadaveris	
Clostridium clostridioforme	
Clostridium difficile	
Clostridium glycolicum	
Clostridium novyi type A	
Clostridium paraputrificum	
Clostridium perfringens	(2)
Clostridium septicum	
Clostridium sordellii	
Clostridium sporogenes	
Clostridium tetani	
Clostridium ramosum	
Eggerthella lenta (Eubacterium lentum,)	(2)
Eubacterium aerofaciens	(2)
Peptostreptococcus spp.	(3)
Peptostreptococcus anaerobius	(2)
Peptostreptococcus asaccharolyticus	(2)
Peptostreptococcus micros	(2)
Peptostreptococcus magnus	(2)
Peptostreptococcus productus	(2)
Propionibacterium acnes	(2)
Staphylococcus saccharolyticus	
Streptococcus mutans	

Results and Discussion

The sampling procedure chosen for the trial was deliberately stringent as it included a dilution step which would not normally be part of the recovery of a strain from cryogenic storage. Survival and recovery of fastidious anaerobes with this protocol, therefore, is a more rigorous test of the system, and makes the results more meaningful.

The overall performance of the Microbank® preservation system for anaerobes was highly satisfactory. Although variations in recovery are apparent between samples, these are probably due to a combination of heterogeneity of inoculum and sampling error. There was no evidence of a gradual decline in recovery over time as compared to the control.

Three organisms failed to survive the trial period; these were Actinomyces odontolyticus, Actinomyces israelii and Prevotella intermedia. The latter two also failed in the control vial, however, and the former was contaminated with a P. acnes, presumably at the date of freezing.

In light of these results the Anaerobe Reference Unit has adopted the Microbank® system for the preservation of strains in its culture collection.

Anaerobe Reference Unit Microbank® Storage Trial 10 Years

Dr J. Brazier, Dr V. Hall.
Anaerobic Reference Centre, Cardiff, UK.

Methods and Materials

Microbank® vials were inoculated with clinical isolates of obligatory anaerobic bacteria referred to the PHLS Anaerobe Reference Unit (now the National Public Health Service for Wales Anaerobe Reference Laboratory, ARL) for confirmation of identity, and were frozen at -80°C for ten years. Isolates for study (n=100) were selected to represent the range of species commonly isolated from clinical material. One bead from each vial had been removed to demonstrate viability after five years and seven years. Beads may have been removed on other occasions.

One bead from each vial was placed aseptically onto Anaerobe Basal Agar (Oxoid, Basingstoke) containing 5% horse blood. Plates were spread for single colonies and were incubated promptly at 37°C in an anaerobic chamber (Concept Plus) for 48 hours. On removal from the chamber, plates were examined for growth of colonies characteristic of the intended isolates. If growth was not apparent at this stage, plates were re-incubated for a further three days.

Results

All 100 strains of anaerobic bacteria of clinical origin were viable after storage in Microbank® vials at -80°C for ten years. Most strains yielded moderate to heavy growth from a single bead after 48 hours incubation but three strains yielded only light growth after five days' incubation. These comprised one of the two strains of Prevotella denticola examined, one of four Fusobacterium nucleatum strains and one strain of Fusobacterium varium.

One hundred anaerobic bacteria representing a wide range of genera isolated from clinical sources remained viable after ten years' storage in Microbank® vials at -80°C. The same 100 vials were previously sampled for viability after five and seven years' storage. However, 34 of those 100 isolates have changed names since their original identification at the ARL. These changes reflect advances in taxonomy and identification methods over the decade.

Some species have simply been placed in novel genera and, in some cases, have changed gender in the process e.g. Eubacterium lentum became Eggerthella lenta. The Gram-positive anaerobic cocci have undergone major taxonomic review resulting in the removal of most former Peptostreptococcus species to novel genera e.g. Finegoldia, Anaerococcus, Peptoniphilus. Additionally, several novel species of anaerobic cocci have been described e.g. Peptoniphilus harei, Peptoniphilus ivorii. Several novel species have been described in other genera e.g. Prevotella, Porphyromonas, Actinomyces. The identification methods used at the ARL have been revised to accommodate such taxonomic changes.

The ARL has developed novel molecular methods, principally amplified 16S ribosomal DNA restriction analysis (ARDRA) for identification of Bacteroides, Prevotella and Porphyromonas and for Actinomyces and other non-sporing Gram-positive bacilli. ARDRA is more accurate and discriminatory than conventional phenotypic tests for identification of these groups; consequently some strains examined in the Microbank® storage trial have been re-designated as a result of retrospective identification by ARDRA. Application of ARDRA to isolates stored at the ARL has resulted in the recognition of several novel Actinomyces species e.g. Actinomyces cardiffensis and a novel genus and species Varibaculum cambriense. By chance, two strains previously included in the Microbank® storage trial as Actinomyces species have subsequently been identified as members of these novel species.

Conclusion

Microbank® vials are easy to use, compact, maintain viability and, therefore, are convenient for the long-term storage of anaerobic bacteria. The vials are particularly practical for repeated retrieval of strains as they ensure that the same strain is sub-cultured on each occasion with minimal effort. The ARL holds a collection of approximately 20,000 referred isolates dating back to the early 1980s. Isolates received in the past decade have been stored in Microbank® vials. This collection is a valuable resource for retrospective research in fields such as evaluation of novel identification methods, monitoring of antimicrobial susceptibilities and development of molecular typing schemes.

IDENTIFICATION	5YRS	7YRS	10YRS
B. distasonis	+	+	+
B. fragilis	+	+	+
B. splanchnicus	+	+	+
B. splanchnicus	+	+	+
B. thetaiotaomicron	+	+	+
B. thetaiotaomicron	+	+	+

IDENTIFICATION	5YRS	7RS	10YRS
B.thetaiotaomicron	+	+	+
B.uniformis	+	+	+
Bacteroides spp.	+	+	+
C. glycolicum	+	+	+
C.bifermentans	+	+	+
C.butricum/beijerinckii	+	+	+
C.cadaveris	+	+	+
C.clostridioforme	+	+	+
C.difficile	+	+	+
C.novyi type A	+	+	+
C.paraputrificum	+	+	+
C.perfringens	+	+	+
C.perfringens	+	+	+
C.ramosum	+	+	+
C.septicum	+	+	+
C.sordellii	+	+	+
C.sporogenes	+	+	+
C.tetani	+	+	+
F. necrophorum	+	+	+
F. russii	+	+	+
F. varium	+	+	+/-
Fusobacterium necrophorum	+	+	+

IDENTIFICATION	5YRS	7RS	10YRS
Porph. asaccharolytica	+	+	+
Porph. endodontalis.	+	+	+
Porph. levii	+	+	+
Porph. uenonis	+	+	+
Porphyromonas spp.	+	+	+
Prev. denticola	+	+	+
Prev. denticola	+	+/-	+/-
Prev. melaninogenica	+	+/-	+
Prev. oris	+	+	+
Prevotella spp.	+	+	+
Prevotella spp.	+	+	+
Prevotella spp.	+	+	+
Peptoniphilus harei	+	+	+
Peptoniphilus harei	+	+	+
Peptoniphilus lacrimalis	+	+	+
Peptoniphilus lacrimalis	+	+	+
Peptostreptococcus spp.	+	+	+
Peptostreptococcus spp.	+	+	+
Anaerococcus octavius	+	+	+
Anaerococcus octavius	+	+	+
Ruminococcus productus	+	+	+
Ruminococcus productus	+	+	+

IDENTIFICATION	5YRS	7RS	10YRS
F. naviforme	+	+	+
F. naviforme	+	+	+
F. naviforme	+	+	+
F. necrophorum	+	+	+
F. necrophorum	+	+	+
F. necrophorum	+	+	+
F. nucleatum	+	+	+
F. nucleatum	+	+	+
F. nucleatum	+	+	+
F. nucleatum	+	+	+
F. nucleatum	+	+	+
F. nucleatum	+	+	+/-
F. nucleatum	+	+	+/-

Microbank ® -20°C Reference Study

Professor Valerie Edwards-Jones.
Manchester Metropolitan University.

The Microbank® bead system will be tested over a two-year period for viability of protected bacteria and Candida spp. Organisms will be sub-cultured and placed into the Microbank® system and stored at -80 and -20°C and revived at set time periods for up to two years.

The details of sub-culture are listed below:

Year 1 1 -12 months / revive monthly
Year 2 12-24 months / revive 3 monthly

The organisms will be revived from beads by dropping one bead into nutrient broth, vortexing and sub-culturing onto the appropriate enriched medium, and incubating in the appropriate atmosphere for a 24/48hr period. Growth or No Growth will be recorded.

Organisms to be tested:

ORGANISM	REFERENCE
Aeromonas hydrophila	NCTC® 8049
Aspergillus niger	NCPF® 2275
Bacillus cereus	NCTC® 7464
Bacillus subtilis	NCTC®10400
Bacteroides fragilis	NCTC® 9343
Burkholderia cepacia	NCTC® 10661
Campylobacter jejuni	NCTC® 11322
Candida albicans	NCPF® 3179
Candida albicans	NCPF® 3255
Clostridium perfringens	NCTC® 8237
Clostridium sporogenes	NCTC® 532
Enterococcus faecalis	NCTC® 775
Enterococcus faecalis	NCTC®12697

ORGANISM	REFERENCE
Escherichia coli	NCTC® 12241
Escherichia coli	NCTC® 11954
Escherichia coli	NCTC® 11560
Escherichia coli O157*	NCTC® 12900
Haemophilus influenzae	NCTC® 12699
Haemophilus influenzae	NCTC® 11931
Klebsiella pneumoniae	NCTC® 9633
Klebsiella pneumoniae**	NCTC® 13368
Listeria monocytogenes	NCTC® 11994
Neisseria gonorrhoeae	NCTC® 8375
Neisseria gonorrhoeae	NCTC® 12700
Proteus mirabilis	NCTC® 10975
Proteus vulgaris	NCTC® 4175
Pseudomonas aeruginosa	NCTC® 12903
Pseudomonas aeruginosa	NCTC® 10662
Salmonella poona	NCTC® 4840
Salmonella typhimurium	NCTC® 12023
Serratia marcescens	NCTC® 13382
Shigella sonnei	NCTC® 8574
Staphylococcus aureus	NCTC® 12981
Staphylococcus aureus	NCTC® 12973
Staphylococcus aureus	NCTC® 12493

ORGANISM	REFERENCE
Staphylococcus aureus	NCTC® 6571
Staphylococcus epidermidis	NCTC® 13360
Streptococcus agalactiae	NCTC® 8181
Streptococcus pneumoniae	NCTC® 12977
Streptococcus pyogenes	NCTC® 12696
Yersinia enterocolitica	NCTC® 12982

* Non-toxigenic Strain, Escherichia coli O157 NCTC® 12900

** ESBL (Extended-Spectrum Beta Lactamases) Klebsiella pneumoniae NCTC® 13368

Results

All strains were viable after storage in Microbank® vials at -20°C and -80°C at 24 months. All strains yielded moderate to heavy growth from a single bead after 24hrs incubation.

Conclusion

Microbank® vials are easy to use, compact, maintain viability and therefore are convenient and effective for the long-term storage of bacteria. The vials are particularly practical for repeated retrieval of strains as they ensure that the same strain is sub-cultured on each occasion with minimal effort.

Long-term storage of multiple large research-led culture

Williams N.J. et al.
Department of Epidemiology and Population Health, Institute for Infection and Global Health, Leahurst Campus, Neston, CH64 7TE.

Many large research programmes have been undertaken at the University of Liverpool involving extensive field sampling of livestock, wildlife, their environment and companion animals. Considerable expense, time and effort are employed to obtain such samples and detect the presence of enteric zoonotic pathogens, namely Campylobacter species, E. coli and Salmonella, and explore antimicrobial resistance among these organisms, as well as commensal E. coli and Staphylococcus species. These collections are held on Microbank® beads (Pro-Lab Diagnostics), which have been in use within our laboratories for the long-term storage (at -70°C) of bacteria for the past 16 years. These culture collections serve as a valuable resource for further examination of the population structure, for example of different bacteria among different hosts in different environments, or to explore specific phenotypes relating to their survival in different environments, therefore it is vital they remain viable for future work.

Within our culture collection we house approximately 65,000 bacterial isolates comprising isolates from previous studies. Some date back to 2000 with isolates of E. coli, C. jejuni, and Staphylococcus species still being resuscitated on a regular basis and used for further studies, both at the University of Liverpool as well as other institutions within the UK. In most cases isolates can be resuscitated easily on appropriate solid media, however for Campylobacter species specifically when kept for a significant number of years (>5 years), we have found from experience that a pre-enrichment step in a non-selective broth incubated overnight under microaerobic conditions before plating onto a non-selective blood agar plate enhances recovery.

The published papers below relate to the most recent studies which we have conducted using Microbank® vials for storage of isolates.

Wedley A, Dawson S, Maddox TW, Coyne KP, Pinchbeck GL, Clegg P, Jamrozy D, Fielder MD, Donovan D, Nuttall T, Williams NJ. (2014). Carriage of Staphylococcus species in the veterinary visiting dog population in mainland UK: molecular characterisation of resistance and virulence. Vet. Microbiol. 170 (1-2):81-8.

Humphrey S, Chaloner G, Kemmett K, Davidson N, Williams N, Kipar A, Humphrey T, Wigley P. (2014). Campylobacter jejuni is not merely a commensal in commercial broiler chickens and affects bird welfare. mBio. 1:5(4):e01364-14.

Merga JY, Royden A, Pandey AK, Williams NJ (2014). Arcobacter spp. Isolated from untreated domestic effluent. Lett.Appl.Microbiol. 59(1):122-6.

Schmidt VM, Williams NJ, Pinchbeck G, Corless CE. Shaw S, McEwan N, Dawson S, Nuttall T. (2014). Antimicrobial resistance and characterisation of staphylococci isolated from healthy Labrador retrievers in the United Kingdom. BMC.Vet.Res. 10 (7). doi: 10.1186/1746-6148-10-17.

Kemmett K, Williams NJ, Chaloner G, Humphrey S, Wigley P, Humphrey T. (2014). The contribution of systemic Escherichia coli infection to the early mortalities of commercial broiler chickens. Avian.Pathol. 43(1):37-42.

Merga JY, Winstanley C, Williams NJ, Yee E, Miller WG. (2013). Complete genome sequence of the Arcobacter butzleri cattle isolate 7h1h. Genome.Announ.1(4): pii:e00655-13.

Merga JY, Williams NJ, Miller WG, Leatherbarrow AJ, Bennett M, Hall N, Ashelford KE, Winstanley C. (2013). Exploring the diversity of Arcobacter butzleri from cattle in the UK using MLST and whole genome sequencing. PLoSOne. 8(2):e55240

Kemmett K, Humphrey T, Rushton S, Close A, Wigley P, Williams NJ. (2013) A Longitudinal Study Simultaneously Exploring The Carriage of APEC Virulence Associated Genes and the Molecular Epidemiology of Faecal and Systemic E. coli in Commercial Broiler Chickens. PLoSOne. 25; 8(6):e67749.

Williams A, Christley RM, McKane SA, Roberts VLH, Clegg PD, Williams, NJ. (2013). Antibiotic resistance changes in equine enteric E. coli during hospitalisation. Vet.J. 196(1):121-6

Parsons BN, Porter CJ, Stavisky JH, Williams NJ, Birtles RJ, Miller WG, Hart CA, Gaskell RM, Dawson S. (2012). Multilocus sequence typing of human and canine Campylobacter upsaliensis isolates. Vet.Microbiol. 153(3-4):391-7.

Ahmed MO, Williams NJ, Clegg PD, van Velkinburgh JC, Baptiste KE, Bennett M. (2012). Analysis of risk factors associated with antibiotic resistant Escherichia coli. Microb.Drug.Resist. 18(2): 161-8.

Maddox T, Clegg PD, Wedley A, Dawson S, Pinchbeck G, Williams NJ. (2012) Cross-sectional Study of Antimicrobial Resistant Bacteria in Horses. Part 2: Risk Factors for Faecal Carriage of Antimicrobial-Resistance Escherichia coli in Horses. Equine Vet J. 44(3):297-303

Wedley AL, Maddox TW, Westgarth C, Coyne KP, Pinchbeck GL, Williams NJ, Dawson S. (2011). Prevalence of antimicrobial-resistant Escherichia coli in dogs in a cross-sectional, community-based study. Vet.Rec. 168(13):354.

Long-Term Storage of Fastidious Campylobacter and Helicobacter using Microbank®

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Abstracts presented at the VIIth International Workshop on Campylobacter, Helicobacter and related organisms, Brussels, Belgium, Sept. 21-25 1993

Long-term storage of fastidious Campylobacter and Helicobacter has proved to be difficult, as various workers have described low recovery rates on revitalising freeze-dried cultures and other methods of preservation. We have used the Microbank® (Pro-Lab Diagnostics, Texas, USA) system, in which porous beads act as carriers to support microorganisms.

Campylobacter mucosalis, C. concisus, C. hyointestinalis, C. curvus, H. pylori and H. fennelliae were tested in this system. Two-or three-day cultures grown on tryptose blood agar plates (Oxoid CM233) under 11' enriched microaerophilic conditions were inoculated into the Microbank® media and then stored at -70°C.

For revitalisation, one bead was placed on the surface of a blood agar plate, allowed to thaw and gently rolled over the surface. Plates were incubated in an enriched microaerophilic environment at 37°C for up to 7 days.

Twenty-two clinical and reference cultures of C. mucosalis and C. conscisus were successfully revitalised from Microbank® storage at 3 weeks, 4 months and 6 months. Five isolates of C. hyointestinalis were successfully revitalised after 7 months. The type strain of C. curvus, NCTC® 11649 was revitalised at 3 weeks, 3 months and 6 months.

Thirty nine of 41 (95%) clinical and reference isolates of H. pylori were revitalised at 2 weeks, 2 months and 8 months. Concurrent revitalisation of freeze-dried cultures of these H. pylori isolates indicated that only 10 of 31 (32%) strains were viable.

Seventeen of 24 (71%) H. fennelliae clinical and reference cultures were viable. Fourteen isolates were revitalised after 6 months, and 3 isolates after 17 to 21 months. The initial inocula must be heavy, and revitalisation should be in a hydrogen-enriched microareophilic atmosphere. The loss of viability in some of the H. pylori and H. fennelliae isolates is attributed to too sparse an inoculum.

The Microbank® system provides a very simple solution to long-term storage of fastidious Campylobacter and Helicobacter strains. As this commercial preservation system is freely available, it obviates the need for specialised media and procedures. Additional testing is required for longer-term storage in the Microbank® system of these medically important microorganisms.

Long-Term Storage of Salmonella Isolates Submitted to the Salmonella Reference Typing Laboratory using Microbank®

The following collection of 312 reference Salmonella strains have been successfully stored on Microbank® for a minimum of 14 years at the Salmonella Reference Typing Laboratory, Veterinary Laboratory Agency, Weybridge, Surrey, UK.

VIAL	SPECIES	VIAL	SPECIES
1	S. Aarhus	25	S. babelsberg
2	S. aberdeen	26	S. bahrenfeld
3	S. abortusovis	27	S. bareilly
4	S. aderike	28	S. barranquilla
5	S. aesch	29	S. bergen
6	S. agona	30	S. berkeley
7	S. agoueve	31	S. berlin
8	S. ajiobo	32	S. berta
9	S. altendor	33	S. bilthoven
10	S. altona	34	S. bispebjerg
11	S. amager	32	S. berta
12	S. amherstiana	33	S. bilthoven
13	S. anatum	34	S. bispebjerg
14	S. anfo	35	S. blegdam
15	S. angoda	36	S. boecker
16	S. ank	37	S. bonariensis
17	S. antarctica	38	S. bonn
18	S. apapa	39	S. borreze
19	S. arechavaleta	40	S. bradford
20	S. arizona	41	S. brancaster
21	S. Arkansas	42	S. bredeney
22	S. artis	43	S. broughton
23	S. ashanti	44	S. bruck
24	S. Askraal	45	S. budapest

VIAL	SPECIES
46	S. bulawayo
47	S. bulovka
48	S. c.suis v. kunzendorf
49	S. california
50	S. canastel
51	S. cannstatt
52	S. caracus
53	S. carrau
54	S. chailey
55	S. chameleon
56	S. chandans
57	S. chingola
58	S. chittagong
59	S. choleraesuis
60	S. chomedey
61	S. christiansborg
62	S. claiborne
63	S. clovelly
64	S. coein
65	S. congo
66	S. cotham
67	S. crossness
68	S. cubana
69	S. dabou
70	S. dahlem
71	S. dakar
72	S. dar es salaam
73	S. derby

VIAL	SPECIES
74	S. deversoir
75	S. djakarta
76	S. dublin
77	S. duisburg
78	S. durban
79	S. durham
80	S. ealing
81	S. eimsbuettel
82	S. eingedi
83	S. elizabethville
84	S. enteritidis
85	S. essen
86	S. etterbeck
87	S. eves
88	S. falkensee
89	S. fischerkietz
90	S. fluntern
91	S. foulpointe
92	S. frankfurt
93	S. freemantle
94	S. freetown
95	S. freiburg
96	S. fresno
97	S. friedenau
98	S. gallinarum
99	S. georgia
100	S. gera
101	S. give

VIAL	SPECIES
102	S. glostrup
103	S. gnesta
104	S. goerlitz
105	S. goldcoast
106	S. haduna
107	S. haelsingborn
108	S. haifa
109	S. halmstad
110	S. harburg
111	S. hartford
112	S. hato
113	S. havana
114	S. heidelberg
115	S. helsinki
116	S. hemingford
117	S. hindmarsh
118	S. hithergreen
119	S. hofit
120	S. houten
121	S. idikan
122	S. illinois
123	S. indiana
124	S. infantis
125	S. inganda
126	S. ipswich
127	S. isangi
128	S. ituri
129	S. jangwani

VIAL	SPECIES
130	S. java
131	S. javiana
132	S. jerusalem
133	S. johannesburg
134	S. kalamu
135	S. kaolak
136	S. kapemba
137	S. karachi
138	S. karamoja
139	S. kegougou
140	S. kentucky
141	S. kiambu
142	S. kibi
143	S. kibusi
144	S. kidderminster
145	S. kiel
146	S. kimuenza
147	S. kirkee
148	S. kisarawe
149	S. koessen
150	S. kokomlemle
151	S. kottbus
152	S. krefeld
153	S. kuessel
154	S. landau
155	S. lanka
156	S. lansing

VIAL	SPECIES
157	S. leiden
158	S. lexington
159	S. lille
160	S. litchfield
161	S. livingstone
162	S. llandoff
163	S. llobregat
164	S. london
165	S. luckenwalde
166	S. luke
167	S. luton
168	S. madjorio
169	S. makiso
170	S. malsatt
171	S. manchester
172	S. manhattan
173	S. manila
174	S. mara
175	S. marembe
176	S. marina
177	S. marshall
178	S. massenya
179	S. matopeni
180	S. mbandaka
181	S. meleagridis
182	S. mgulani
183	S. midway

VIAL	SPECIES
184	S. mikawasima
185	S. millesi
186	S. milwaukee
187	S. minneapolis
188	S. minnesota
189	S. mississippi
190	S. mobeni
191	S. moero
192	S. molade
193	S. mondeor
194	S. mons
195	S. monschaui
196	S. montevideo
197	S. morehead
198	S. morningside
199	S. mountpleasant
200	S. mpouto
201	S. muenster
202	S. naestved
203	S. nagoya
204	S. napoli
205	S. nchanga
206	S. nessziona
207	S. newbrunswick
208	S. newport
209	S. newyork
210	S. nigeria

VIAL	SPECIES	VIAL	SPECIES
211	S. nima	238	S. portsmouth
212	S. nitra	239	S. pullorum
213	S. nottingham	240	S. putten
214	S. noya	241	S. rawash
215	S. nyanza	242	S. rideau
216	S. oakland	243	S. riggil
217	S. ochiogo	244	S. riogrande
218	S. offa	245	S. rosenthal
219	S. ohio	246	S. rubislaw
220	S. okerara	247	S. ruiru
221	S. omifisan	248	S. rumford
222	S. ona	249	S. saintpaul
223	S. onderstepoort	250	S. salford
224	S. oranienburg	251	S. sarajone
225	S. ordonez	252	S. schlessheim
226	S. orientalis	253	S. selandia
227	S. oslo	254	S. sendai(miami)
228	S. ouakam	255	S. senegal
229	S. panama	256	S. senftenberg
230	S. papuana	257	S. seremban
231	S. Paratyphi A	258	S. shubra
232	S. Paratyphi B	259	S. simsbury
233	S. patience	260	S. singapore
234	S. pietersburg	261	S. sinthia
235	S. plymouth	262	S. soesterberg
236	S. pomona	263	S. solt
237	S. poona	264	S. stanleyville

291289VIAL	SPECIES
265	S. sternschanze
266	S. stockholm
267	S. stourbridge
268	S. strasbourg
269	S. sundsvall
270	S. tamale
271	S. tamberma
272	S. taunton
273	S. tees
274	S. telaviv
275	S. telelkebir
276	S. telhashomer
277	S. teltow
278	S. thielallee
279	S. thomasville
280	S. thompson
281	S. thompson
282	S. tione
283	S. toricada
284	S. toucra
285	S. typhi
286	S. typhimurium
287	S. uccle
288	S. uzaramo
289	S. vejle
290	S. vinohrady
291	S. virchow

VIAL	SPECIES
292	S. vitkin
293	S. vleuten
294	S. vogan
295	S. wa
296	S. wandsbek
297	S. wandsworth
298	S. warral
299	S. wassenaar
300	S. wayne
301	S. weslaco
302	S. westerstede
303	S. westhampton
304	S. widemarsh
305	S. wildwood
306	S. windermere
307	S. worthington
308	S. wuppertal
309	S. wyldergreen
310	S. yeerongpilly
311	S. yerba
312	S. yoruba

Cryopreservation of Fungal Spores using Microbank®

Dr Chandler
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A simple method is described for the cryopreservation of conidia, stored adhering to small porous beads in a robust polypropylene vial. For recovery, a single bead is removed from the vial and streaked onto a solid growth medium. Preparative work is minimised so the method is rapid. The storage life of an isolate can be increased greatly if a 'seed lot' system is employed.

The most reliable way to store fungi for extended periods is in liquid nitrogen or in the vapour of liquid nitrogen. Elliott (1976) used polypropylene drinking straws to store strains of Agaricus bisporus in liquid nitrogen as the straws were safer and less expensive than using glass ampoules. The small size of the straws also permitted greater storage capacity and sample replication in vivostats. However, there are problems with drinking straws as their small size makes them difficult to handle, they are hard to check for leaks after sealing and they can burst when removed from a vivostat. The preparative work is also time-consuming, so cultures tend to be handled in batches which increases the chances of cross contamination. Stalpers, De Hoog and Vlug (1987) recommended the use of special apparatus for large-scale application of the straw technique but this apparatus is not available commercially.

This note describes cryopreservation of conidia of mitotic, entomopathogenic fungi on porous beads. The Microbank® storage system (Pro-Lab Diagnostics) is a commercial variant of a system designed originally for storing bacteria (Feltham et al, 1978) but also appears suitable for storing certain fungi. In our laboratory at Wellesbourne, cultures are stored in the vapour above liquid nitrogen, but porous beads could be used for storing microorganisms in a deep freeze (ca -70°C). The Microbank® system consists of sterile vials containing beads (3 mm diam.) which act as carriers to support the microorganisms. Each vial has a volume of 2 ml and contains 25 beads in 1 ml of a cryopreservative (usually 10% or 15% glycerol). As the Microbank®'system is available commercially, preparative work is kept to a minimum. Vials and beads are available in a range of colours to aid identification and vials are robust and easy to handle.

In this laboratory, the preparation, inoculation and recovery of conidia are performed in a microbiological safety cabinet. A suspension of conidia, prepared in sterile 0.05% Tritron X-100 surfactant, is placed in a sterile Eppendorf tube, centrifuged, washed and centrifuged again. The pellet is then resuspended in cryopreservative taken from one of the vials and pipetted back into that vial, which is inverted 3 to 4 times to allow conidia to adhere to the beads. Most of the cryopreservative is then removed to prevent the beads sticking together during freezing, but a thin layer of free liquid can be left at the bottom of the vial to allow recovery of the fungus should the conidia fail to adhere to the beads. Vials are frozen overnight in a deep-freeze at -70° and transferred to a vivostat the following morning. The vials are stored in the vapour above liquid nitrogen as this avoids the problem of liquid nitrogen entering the vials through cracks or leaks.

For recovery, vials are removed from the vivostat and one bead is removed with sterilised forceps and streaked, using a loop, onto the surface of a suitable solid medium. Vials are prevented from thawing during this procedure by placing them into an insulated block of aluminium (10 x 8 x 4 cm) or 'Cryoblock', which has sample wells drilled into it (PL.155 available from Pro-Lab Diagnostics). The block is stored in the deep-freeze and can be cooled further in liquid nitrogen immediately before use. The method is simple and rapid so that the vials are out of the vivostat for only two to three minutes. The temperature of a vial placed within the cryoblock was measured on the laboratory bench at room temperature (approximately 20°C) using a thermocouple, after they had been frozen overnight at -70°C. Consistently, the cryoblock kept the vial at a temperature below -60°C for five min. below -50°C for 15 min and below -40°C for 30 min.

The method has been used to store approximately 50 isolates of Beauveria bassiana, Metarhizium anisopliae, Verticillium lecanii and Paecilomyces spp. Prima facie, the method appears to be suitable only for storing conidia: it remains to be seen whether it is also suitable for preserving other life stages, e.g. hyphal bodies produced in liquid culture. To date, Conidia have been stored for 18 months, with recovery of a bead every 3 months, so that the suitability of the method for very long-term preservation has not been assessed. However, all isolates stored in this way have been recovered without contamination and loss of pathogenicity to target insects has not been observed in our routine bioassays. A preliminary assessment of the germination of conidia was performed for two isolates each of B. bassiana, M. anisopliae and V. lecanii.

Each isolate, obtained from cultures stored previously for a minimum of six years in polypropylene straws under liquid nitrogen, had been stored for 18 months using the bead system. Conidia were washed from beads in 1 ml 0.05% Triton X-100 and an aliquot (0.1 ml) pipetted onto Sabouraud dextrose agar in 5.5 cm Petri dishes. Dishes were incubated at 23°C for 24 hr, after which Conidia were stained with lactophenol cotton blue and germinated/ ungerminated conidia were counted using a compound miscroscope. Conidia were considered to have germinated when the length of the germ tube exceeded the width of the conidium. Three replicates were used with a minimum of 300 conidia counted each time. Germination of conidia was greater than 95% in all cases. Each bead held between 105 and 106 conidia.

An isolate can be stored in a 'seed lot' system to increase its storage life. Two vials are prepared from the isolate. One vial, the 'working' vial, is used for preparing cultures for experimental work. When all the beads from the working vial have been used, a bead from the other 'seed' vial is removed, a culture is grown from it and is used to prepare a second working vial. In this laboratory, a further level of storage has been added. A slope culture from the working vial is kept in a refrigerator at 5°C for 3 months and cultures for experiments are grown from this. As each working vial contains 25 beads, it should last for over 6 years. Theoretically, therefore, an isolate stored using this method of cryopreservation can be accessed for 156 years before it needs to be replaced.
This work was supported by the Ministry of Agriculture, Fisheries & Food.

References

Elliott, T.J. (1976). Alternative ampoule for storing fungal cultures in liquid nitrogen. Transactions of the British Mycological Society 67. 545-546.

Feltham, R.K.A, Power A.K., Pell P.A. & Sneath, P.H.A. (1978). A simple method for storage of bacteria at -76° C. Journal of Applied Bacteriology 44. 313-316.

Stalpers, J.A., De Hoog A. & Vkyg IJ. (1987). Improvement of the straw technique for the preservation of fungi in liquid nitrogen. Mycologia 79. 82-89.

Storage of NCTC Reference Set of Cultures Selected for Quality Control & Laboratory Accreditation using Microbank®

Terence J Donovan PhD.

Environmental Microbiologist, Ashford Public Health Laboratory

A simple method is described for the cryopreservation of conidia, stored adhering to small porous beads in a robust polypropylene vial. For recovery, a single bead is removed from the vial and streaked onto a solid growth medium. Preparative work is minimised so the method is rapid. The storage life of an isolate can be increased greatly if a 'seed lot' system is employed.

Introduction

Ashford PHL carried out an investigation using cultures supplied by NCTC® to select cultures suitable for use in QC procedures in Food, Water and Dairy products testing. These were initially directed to Public Health Laboratory's requirements but were extended to cover other laboratories' requirements.

Following feedback from colleagues, both in the PHLS and other sources, a set of 44 cultures were selected.

Method

The freeze-dried vials received from NCTC® were checked for vacuum integrity using a spark tester. Vials were then opened following the NCTC® instructions under a Class III safety cabinet. The hydrated cultures were then inoculated into appropriate media (in most cases Blood Agar). The incubation temperature, gaseous requirement and length of incubation were selected as appropriate to the culture eg 37°C aerobic 24 hours for most cultures.

The incubated cultures were examined for viability and purity. A subculture was prepared for identification check as appropriate to the individual culture.

Inoculation of the Vials and Beads in the Laboratory

Using the primary culture obtained directly from the NCTC® vial the vials were inoculated following the Pro-Lab Diagnostics Procedure A Preparation.

Storage

The vials were placed in a -70°C cabinet (Kelvinator).

Check of Viability of Beads

All inoculated beads were checked for viability within 1 week of inoculation; all were viable.

Recovery Method

The Pro-Lab Diagnostics Procedure B for recovery was followed. In addition “cold blocks” were used to carry the culture from the -70°C cabinet and during testing. Recovery was limited to 6 cultures at a time to reduce temperature loss. The beads were inoculated onto the appropriate media and appropriate culture conditions were used. Recovery was assessed by a semi-quantitative scoring method.

Recovery Results

The dates of bead inoculation and last date of viability (recovery) check are given on the enclosed list.

All cultures were viable at the recent dates shown. The majority have also been tested previously.

All cultures remained viable with no quantitative loss of viability detected. The majority of the cultures have been stored on beads at -70°C for over 2 years with no loss of viability.
Tests for storage at other temperatures e.g. -20°C were not performed.

VIAL	IDENTIFICATION	REFERENCE	MONTHS STORED
1	Zygosaccharomyces rouxii	NCPF® 3879	26
2	Aspergillus niger	NCPF® 2275	26
3	Bacillus subtilis	NCTC® 7464	26
4	Bacillus subtilis	NCTC® 10400	26
5	Campylobacter jejuni (BT1)	NCTC® 11322	26
6	Saccharomyces cerevisiae	NCTC® 3178	26
7	Citrobacter freundii	NCTC® 9750	27
8	Clostridium difficile	NCTC® 11204	26
9	Clostridium perfringens	NCTC® 8237	26
10	Enterobacter aerogenes	NCTC® 10006	27
11	Enterococcus faecalis	NCTC® 775	27

VIAL	IDENTIFICATION	REFERENCE	MONTHS STORED
12	Escherichia coli	NCTC® 10418	9
13	Escherichia coli O157:H7	NCTC® 12079	26
14	Lactobacillus casei	NCTC® 10302	17
15	Legionella pneumophila SG 1	NCTC® 12821	27
16	Listeria monocytogenes 4b	NCTC® 11994	26
17	Proteus mirabilis	NCTC® 10975	26
18	Pseudomonas fluorescens	NCTC® 10038	24
19	Pseudomonas aeruginosa	NCTC® 10662	26
20	Pseudomonas cepacia	NCTC® 10661	26
21	Salmonella Typhimurium	NCTC® 12023	26
22	Shigella sonnei	NCTC® 8574	26
23	Staphylococcus aureus	NCTC® 6571	26
24	Staphylococcus epidermidis	NCTC® 11047	20
25	Lactobacillus lactis	NCTC® 662	26
26	Listeria ivanovii	NCTC® 11846	24
27	Campylobacter coli	NCTC® 11366	25
28	Vibrio furnissii	NCTC® 11218	26
29	Vibrio parahaemolyticus	NCTC® 10885	26
30	Yersinia enterocolitica SG1	NCTC® 10460	26
31	Klebsiella aerogenes	NCTC® 9528	26
32	Aeromonas hydrophila	NCTC® 8049	26
33	Acinetobacter lwoffii	NCTC® 5866	26
34	Serratia marcescens	NCTC® 11935	26

VIAL	IDENTIFICATION	REFERENCE	MONTHS STORED
35	Edwardsiella tarda	NCTC® 11934	26
36	Proteus rettgeri	NCTC® 7475	26
37	Enterobacter cloacae	NCTC® 11936	26
38	Vibrio cholerae Non O:1/O:24	NCTC® 11348	26
39	Salmonella Poona O13, 22	NCTC® 4840	26
40	Rhodococcus equi	NCTC® 1621	26
41	Staphylococcus aureus	NCTC® 1803	26
42	Clostridium bifermentans	NCTC® 506	26
43	Clostridium sporogenes	NCTC® 532	26
44	Listeria innocua	NCTC® 11288	26

Conclusion

Storage of the NCTC® Reference Set of Cultures has been demonstrated under normal working conditions in a Public Health Laboratory using the Microbank® system, manufactured by Pro-Lab Diagnostics.

This method of storage of cultures is recommended for use in microbiological laboratories requiring reference cultures for Laboratory Accreditation and QC procedures. The NCTC® set of cultures is recommended for this purpose.

Storage and Reapeated Rovery of Neisseria gonorrhoeae using Microbank®

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Introduction

Due to the fastidious nature of Neisseria gonorrhoeae a simple, inexpensive and efficient system for the storage and recovery of clinical isolates and quality control strains is required for good clinical laboratory practice, in research and for epidemiological studies. Various methods such as use of cooked meat broth, lyophilisation or freezing in liquid nitrogen are available for the storage of bacteria but none is ideal, particularly for gonococci. The limitations associated with these methods are the variable recovery of bacteria, the time and inconvenience involved in the preparation and maintenance of cultures and the financial cost in the purchase and maintenance of expensive equipment.

The technique of storing organisms at -70°C described by Nagel and Lawrence in 1971 has given rise to simple and convenient commercial storage systems such as the Pro-Lab Microbank® which uses coloured beads in a 'cryovial' containing cryopreservative fluid. After inoculation and storage a single bead can be removed to inoculate culture media.

We examined the Microbank® system with a view to its overall convenience of use for storage and recovery. The recovery rate after medium-term storage and repeated sampling was analysed and an evaluation made of the efficacy of recovery in relation to the spectrum of antigenic types (serovars) of gonococci that occur in nature.

Materials and Methods

Bacterial strains

One hundred clinical isolates of N. gonorrhoeae, including 30 penicillinase-producing N. gonorrhoeae were included in the study. These strains represented the wide variety of antigenic types of gonococci encountered in natural infection and comprised 26 serogroup IA strains covering eight different serovars, 61 serogroup IB strains covering 14 different serovars and 13 serogroup IB strains which were non-typeable with the standard monoclonal antibody serotyping panel.

Preservation and storage of strains

Using a sterile cotton bud, gonococcal colonies were harvested from an 18-24 hour culture on modified New York City medium and a suspension made in the cryopreservative fluid of the cryovial approximately equivalent to a McFarland No. 4 standard. The inoculated vial was closed and the contents inverted 4-5 times to coat the beads with bacteria. Excess cryopreservative fluid was removed with a sterile pastette. The vial was closed and immediately placed in a -70°C freezer.

Retrieval of bacteria

All 100 gonococcal isolates were sampled each month for 24 consecutive months. Twenty cryovials at a time were removed from the -70°C freezer, using an aluminium transfer block to retain a low temperature. Using sterile forceps a single bead was removed from the cryovial, placed onto the surface of a culture place containing modified New York City medium, and allowed to thaw. A sterile loop was used to streak out the area around the bead to obtain separate colonies and the culture plates incubated for 48 hours in a carbon dioxide-enriched atmosphere.

Serotyping

Serotyping of the gonococcal strains was performed using the Genetic Systems 6 panel of monoclonal antibodies.

Statistical analysis

The chi-squared test was used for all statistical analysis.

Results

As shown in Table 1, the overall recovery rate from 2400 retrievals for the 24 months of the trial was 98.6% (2365/2400), and all strains were recovered in eight of the 24 months (100% recovery). The recovery rate for the remaining months ranged from 99% to 96%: 99% in five of the months, 98% in five of the months, 97% in four of the months and 96% in two of the months. Although the recovery rate was extremely good there were 0.8% (9/1191) failures in the first 12 months compared with 2.2% (26/1174) in the last 12 months - a significant difference ($x^2 = 8.4$; $P<0.01$).

Thirteen separate isolates accounted for the 35 (1.5%) failures and the distribution of these failures by serovar and month is shown in Table 2. Five isolates failed on only one occasion, four on two occasions, two isolates on five occasions and two isolates on six occasions. The distribution of the 13 serovar failures in relation to the total number of isolates for each of the 23 sereovars tested is given in Table 3. Although there were only three serovar IB15 isolates, each one failed on at least one occasion and together they accounted for 37.1% (13/35) of the total failures. Serovars IA16 and IB25 were also associated with multiple failures on five or more occasions (Table 2).

Thirty-five isolates comprising serovars IA02, IA06, IB01, IB02 and IB03, classified as major serovars from continuous prevalence studies in our geographical area, accounted for only 2.0% (1/35) of the failures. The remaining 65 isolates, classified as minor serovars, accounted for 18.5% (12/65) of the failures - a significant difference ($x^2 = 4.9$; $P<0.05$).

Table 1: Monthly recovery rate for 100 gonococcal isolates

MONTH	NO. OF RECOVERED STRAINS	NO. OF FAILURES
1	100	0
2	100	0
3	100	0
4	99	1
5	100	0
6	100	0
7	100	0
8	100	0

MONTH	NO. OF RECOVERED STRAINS	NO. OF FAILURES
9	100	1
10	100	2
11	100	2
12	99	3
13	100	1
14	100	3
15	100	3
16	100	2
17	96	4
18	100	0
19	97	3
20	98	2
21	99	1
22	99	1
23	96	4
24	98	2
TOTAL	2365 (98.6%)	35 (1.5%)

Table 2: Distribution of 35 failures by serovar and month

SEROVAR	MONTH
IA05	15, 16
IA16	11, 15, 17, 19, 23
IA21	17
IA25	17, 19
IB02	21
IB15	12, 24
IB15	11, 14, 16, 20, 22
IB15	13, 14, 15, 20, 23, 24
IB19	23
1B25	4, 9, 10, 12, 14, 17
1B29	19
1B00	10, 23
IB00	12

IB00 (Non-typeable strain)

Table 3: Distribution of 13 failures in relation to individual serovars

SEROVAR	NO. OF ISOLATES	NO. OF FAILURES
IA02	6	0
IA04	3	0
IA05	2	1
IA06	5	0
IA07	1	0

SEROVAR	NO. OF ISOLATES	NO. OF FAILURES
IA16	4	1
IA21	3	1
IA25	2	1
IB01	10	0
IB02	7	1
IB03	7	0
IB05	2	0
IB06	8	0
IB07	5	0
IB08	4	0
IB15	3	3
IB17	7	0
IB19	2	1
IB25	2	1
IB26	1	0
IB29	2	1
IB31	1	0
IB00	13	2

IB00 (Non-typeable strain)

Discussion

Lyophilisation has long been accepted as the 'gold standard' method for the preservation of microorganisms, but the high cost in equipment and processing time precludes its use in many routine laboratories. Modern technology has made -70°C facilities readily available and the small capacity required to store large numbers of isolates makes cryovial storage systems extremely convenient for the clinical laboratory which requires easy access to strains. Concerns over refrigeration failure and the subsequent loss of valuable strains can be alleviated by fitting carbon

dioxide back-up systems, designed to activate at a pre-set temperature to the freezer.

Nagel and Lawrence first described a method for the preservation of multiple replicate units of bacteria using sterile glass beads and a mixture of equal parts of broth culture and horse blood allowing storage of at least 200 beads in a plastic tube at -70°C. In a subsequent study Feltham et al. used different concentrations of cryoprotectants in the storage media used to make the bacterial suspensions, and stored the beads at -70°C. They observed a reduction in the number of viable bacteria with nutrient broth containing 15% dimethyl sulphoxide. Nutrient broth supplemented with either: 10% dimethyl sulphoxide, 10% glycerol or 15% glycerol, showed no such reductions. In a further study White and Sand demonstrated the viability of organisms after storage at -76°C for two years, using glass beads and brain-heart infusion broth containing 10% glycerol as the emulsifying fluid.

In this study we have shown that the Microbank® system offers a simple commercially available system for medium-term storage and multiple recovery of N. gonorrhoeae. The overall recovery rate of 98.6% is extremely good and, together with the ability to sample up to 25 times, represents substantial cost benefits. The failure of four isolates to grow at least five times each may be associated with the strains, or may be a simple physical problem of insufficient primary inoculum in these vials. The overall recovery rate could possibly be improved with the use of special recovery medium. Morton and Smith advocated the use of a solution of 20% sucrose in phosphate-buffered saline for the recovery of fastidious organisms such as Neisseria sp, though clearly this is not essential for the vast majority of gonococcal isolates.

There was a small but significant decrease in recovery in the last 12 months of the trial, which may reflect a sampling problem rather than a temporal phenomenon and further long-term studies are underway to differentiate between these possibilities. The finding that failures were significantly associated with minor serovars suggests that the transmission/viability of minor serovars may be lower than that of common serovars and could be a significant factor in the overall epidemiology of gonococcal infection. The selective loss of minor serovars on storage could also lead to a bias in epidemiological studies based on isolates that have been stored for some time.

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Validation for Cryostorage of Brucella spp. using Microbank®

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The Brucella research section of the Veterinary Laboratories Agency (VLA) under the auspices of the Food and Agriculture Organisation (FAO)/World Health Organisation (WHO) Collaborating Centre for Reference and Research on Brucellosis and on behalf of the Office International des Epizooties (OIE) Brucellosis Reference Centre, has as one of our roles to advise other laboratories on the identification and preservation of Brucella strains. As finances for these laboratories can be a problem it is not always possible to freeze-dry samples. On a recent VLA workshop in Morocco the delegates from North African countries were presented with Microbank® cryovials as an alternative storage method. They thought that these were ideal for their needs. We would like to be in a position to recommend these vials to colleagues at other laboratories worldwide with confidence of their suitability for the long-term storage of Brucella.

Materials and Methods

A representative isolate of each sub-species and in addition the most fastidious strain of Brucella abortus biovar 2 (Advances in Brucellosis Research, Texas A & M University Press, Texas) henceforth referred to as a set, were sub-cultured onto serum dextrose agar (SDA) to check for purity. Each reference strain was freeze-dried allowing one vial for each date of testing in the trial; they were also added to the Microbank® cryovials in accordance with the manufacturers instructions. In order to ascertain the effects of freeze/thawing an individual vial was set up for each of the storage conditions and each testing date. After one month of storage at +4°C the first set of freeze dried vials were reconstituted and subbed onto SDA plates to give a circle of approximately 5cm in diameter. The Microbank® cryo vials from the -20°C and -80°C freezers, a bead was removed from these and spread on SDA plates to give a circle approximately 5 cm in diameter. All plates were then incubated at 37°C in a 10% CO2 atmosphere for 4 days. Growth was examined for morphology and the quantity was compared allowing a deviation of approximately 25% growth between the cultures of different storage conditions. Although the inoculum was not standardised, it is important that sufficient quantity of the isolate remains viable in order to carry out further work on the isolates. The morphology was examined visually, aided using obliquely reflected light from under the culture.

After six months this was repeated, however this time beads were removed from the original Microbank® cryovials, opened at one month and an additional new bead from a fresh vial was removed for the sixth month stage in order to assess the effect of freeze-thawing. This process was also repeated after one year of storage.

Results

Table Key

++	Growth comparable to freeze-dried vials
+-	Growth, but insufficient to be comparable to freeze-drying
+	Freeze-dried vials' growth (used as the standard for comparison against)
-	No growth
?	Change in morphology of the culture
!	Contaminant present

Freeze-dried vials stored at +4°C

ISOLATE	1 MONTH	6 MONTHS	1 YEAR
Brucella melitensis (biovar 1) 16M	+	+	+
Brucella abortus (biovar 1) 544	+	+	+
Brucella suis (biovar 1) 1330	+	+	+
Brucella canis RM6/66	+	+	+
Brucella neotomae 5K33	+	+	+
Brucella abortus (biovar 2) 86/8/59	+	+	+
Brucella ovis 63/290	+	+	+

Microbank® cryovials stored at -20°C

ISOLATE	1 MONTH	6 MONTHS	1 YEAR	TIME IN STORAGE
Brucella melitensis	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella abortus (biovar 1) 544	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year

ISOLATE	1 MONTH	6 MONTHS	1 YEAR	TIME IN STORAGE
Brucella suis (biovar 1) 1330	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella canis RM6/66	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella neotomae 5K33	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella abortus (biovar 2) 86/8/59	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella ovis 63/290	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year

Microbank® cryovials stored at -80°C

ISOLATE	1 MONTH	6 MONTHS	1 YEAR	TIME IN STORAGE
Brucella melitensis (biovar 1) 16M	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella abortus (biovar 1) 544	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella suis (biovar 1) 1330	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella canis RM6/66	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella neotomae 5K33	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella abortus (biovar 2) 86/8/59	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year
Brucella ovis 63/290	++ ++ ++	++ ++ ++	++ ++ ++	1 Month 6 Months 1 Year

Conclusion

The preliminary results have so far shown that the procedure could be a cost-effective alternative to freeze-drying negating the problems associated with freeze storage of cultures, such as loss of viable organisms through lysis during freeze-thawing owing to the formation of ice crystals. The yield and morphology of the Microbank® cryovials in this first year have proved to be comparable with that of freeze-drying. All the cultures were still viable after the first year of this trial and as of yet there has been no significant difference in the products stored at -20°C and those stored at -80°C.

This study has also shown that the freeze-thawing of the Microbank® cryovials has limited effect so far, proving the efficacy of the cryopreservative.

Long-Term Preservation of Fungal Isolates in Microbank® Vials

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Since 1994, 198 yeasts and 391 moulds belonging to 25 and 37 species, respectively, were stored in Microbank® cryogenic vials at < -130°C in liquid nitrogen and at -70°C in a freezer. All of the isolates, with the exception of 45 yeasts and 15 dermatophytes, were recovered from both storage temperatures. Good reproducibility was demonstrated for amphotericin B, fluconazole and voriconazole MICs determined for random isolates.

Long-term preservation of fungal strains is essential for their in-depth study; however, both the viability and the stability of living cells should be ensured during the preservation period. Fungal isolates are usually preserved in water at room temperature, an easy and economical procedure introduced for fungi by Castellani in 1939. Because the stability of fungal cells was not ensured by this simple procedure, other methods have been suggested, such as preservation in soil or on oil or water-covered slants, cryopreservation either in liquid nitrogen or at low temperature (-20 and -70°C), and lyophilisation (the freeze-drying procedure). Cryopreservation in liquid nitrogen and lyophilization are the methods recommended and used by the American Type Culture Collection. Although lyophilisation of living cells provides a mechanism for stabilising these cells for long periods of time, this procedure is cumbersome and lengthy and requires expensive equipment. On the other hand, storage in liquid nitrogen vapour (above the liquid at < -130°C) is a more convenient and less expensive alternative for long-term storage of living cells. Storage above the liquid nitrogen prevents leakage of the liquid nitrogen into the vials.

The Microbank® system (Pro-Lab Diagnostics) consists of sterile vials that contain 25 porous, coloured beads and a cryopreservative fluid; this system was originally developed for storage of bacterial cells. The beads are acid washed, and their porous nature allows the cells to adhere to the bead surface; the beads serve as carriers for the cells being stored (Microbank® package insert). When an isolate is stored in this way, 25 or more identical cultures can be preserved. The purpose of this study was to evaluate the preservation in Microbank® sterile vials of yeast and mould clinical isolates that were received

from 1994 to the end of 2002 at the VCU Medical Center (Richmond, Va.) and the Valme University Hospital (Seville, Spain). Two temperatures (< -130°C {liquid nitrogen vapour} and -70°C {freezer}) were evaluated.

Fresh, pure cultures of 6,198 yeast and yeast-like organisms and 391 moulds (Table 1) were grown on either Sabouraud dextrose agar (for yeasts) or potato dextrose agar (for moulds) at 35°C; some isolates of dermatophytes, Histoplasma capsulatum, Blastomyces dermatitidis and Alternaria spp. were incubated at 30°C. Yeast and yeast-like isolates were incubated for 48 to 72 hours and moulds were incubated for 7 to 15 days. Each isolate was stored in accordance with the directions of the manufacturer. For each isolate, the cryogenic fluid of two Microbank® vials was inoculated with the fungal growth to a density approximately equivalent to a McFarland standard of 3 or 4. The inoculated fluid was mixed four or five times to emulsify the suspension and to bind the cells to the beads. The extraneous cryogenic fluid was then removed, leaving the inoculated beads as free of liquid as possible to prevent the beads from sticking together during freezing but allowing a thin layer of suspension to stay at the bottom of the vial. The vials were then held overnight at -70°C. After overnight freezing, one of the vials was stored in liquid nitrogen vapour (> -130°C) and the other was left at -70°C.
The viability and purity of the strains were monitored immediately after storage, at 1 and 6 months after storage, and once a year subsequently as follows. One of the inoculated beads was removed under aseptic conditions with a sterile needle, and each vial was returned immediately to the corresponding low temperature; the bead was then inoculated onto either Sabouraud dextrose agar or potato dextrose agar for at least 20 days. Both viability and the morphological characteristics of each culture were observed.
Each mould isolate was considered viable if the rate of growth present was the same as that of the original culture and if the morphology and colour of the colony matched the fungal identification documented for each strain. All of the mould strains, with the exception of 15 (of 61) dermatophyte isolates, were recovered each time from both storage temperatures and showed the initial colony characteristics, growth rates, and morphologies (Table 1). Only isolates of B. dermatitidis, H. capsulatum, and Alternaria spp. required more than one bead for harvesting; they required two to four beads. These results are in agreement with those described by Chandler, who preserved 50 uncommon moulds for 18 months in Microbank® vials and found that only one bead was necessary for the recovery of most isolates. Each yeast strain was considered viable if growth was present; the identification and purity of yeasts were also randomly validated on CHROMagar medium. A very small percentage of yeasts (0.7%) were not recovered; Candida dubliniensis had the lowest recovery rate (33%; 28 of the 42 isolates were not recovered). The stability was validated by determining the antifungal susceptibilities of random samples of yeasts (200 isolates) and moulds (50 isolates) stored at both temperatures. Amphotericin B, fluconazole and voriconazole MICs were determined by following NCCLS guidelines (documents M27-A2 {for yeasts} and M38-A {for moulds}) before storage and 6 months and 4 years after preservation. MICs for the isolates after storage were either the same as, or within three dilutions of the MICs before storage, which is the criterion used in NCCLS studies to obtain percentages of intra-and inter-laboratory reproducibilities as well as for establishing quality control MIC ranges.
In general, the effects of both storage temperatures on the stability and viability of stored isolates were similar, which is fortunate because most laboratories have a -70°C freezer.
The advantage of using the Microbank® system over other cryogenic systems is its commercial availability. The time-consuming procedure of preparing other preservative devices such as drinking straws or cryogenic fluid is avoided; Microbank® vials are stored at room temperature prior to use. The harvesting of individual isolates is easier than that described by Pasarell and McGinnis, in which a portion of the frozen culture is chipped and sub-cultured. Because vials should not be outside the low-temperature device for more than 3 minutes to avoid thawing, it is recommended that the frozen vials be placed in an insulated cryoblock during harvesting.
In conclusion, the Microbank® system appears to be an easy, convenient, economical and effective tool for the preservation of fungal isolates other than dermatophyte and C. dubliniensis strains. Longer monitoring of isolates and storage of other species would further validate the reliability of this system for the cryogenic preservation of yeast and mould strains. Also, the stability of fungal cells should be further assessed by molecular parameters.

TABLE 1 - Fungal isolates preserved between 1994 and 2002

SPECIES	NO. OF ISOLATES STORED	NO. (%) OF ISOLATES NOT RECOVERED
Yeasts and yeast-like organisms:	4,453	5
Candida albicans	5	0
Candida ciferri	42	28
Candida dubliniensis	28	0
Candida famata	359	0
Candida glabrata	33	0
Candida guilliermondii	6	0
Candida humicola	10	0
Candida kefyr	279	5
Candida krusei	20	0
Candida lambica	5	0
Candida lipolytica	43	0
Candida lusitaniae	352	3
Candida parapsilosis	3	0
Candida rugosa	401	2
Candida tropicalis	14	0
Candida zeylanoides	112	1
Cryptococcus neoformans	3	0
Cryptococcus albidus	9	0
Cryptococcus laurentii	4	0
Hansenula anomala	1	0

SPECIES	NO. OF ISOLATES STORED	NO. (%) OF ISOLATES NOT RECOVERED
Sporobolomyces salmonicolor	2	0
Trichosporon beigelii	6	1
Rhodotorula rubra	1	0
Rhodotrula glutinis	7	0
Saccharomyces cerevisiae	0	0
TOTAL YEAST AND YEAST-LIKE ORGANISMS:	6,198	45 (0.7)
Moniliaceous moulds:		
Aspergillus fischeri	1	0
Aspergillus flavus	24	0
Aspergillus fumigatus	117	0
Aspergillus niger	10	0
Aspergillus nidulans	9	0
Aspergillus terreus	27	0
Aspergillus sydowii	1	0
Aspergillus versicolor	1	0
Fusarium incarnatum	1	0
Fusarium moniliforme	5	0
Fusarium solani	10	0
Fusarium oxysporum	5	0
Paecilomyces lilacinus	9	0
Rhizopus arrhizus	7	0

SPECIES	NO. OF ISOLATES STORED	NO. (%) OF ISOLATES NOT RECOVERED
Sporobolomyces salmonicolor	2	0
Rhizomucor pusillus	1	0
Trichoderma longibrachiatum	4	0
TOTAL MONILIACEOUS MOULDS:	232	0
Dematiaceous moulds:		
Alternaria spp.	4	0
Bipolaris hawaiiensis	3	0
Bipolaris spicifera	3	0
Cladophialophora bantiana	8	0
Cladosporium cladosporioides	1	0
Curvularia spp.	4	0
Dactylaria gallopava	3	0
Exophiala jeanselmei	6	0
Exophiala spinifera	2	0
Phoma spp.	1	0
Scedosporium apiospermum	17	0
Scedosporium prolificans	10	0
Wangiella dermatitidis	9	0
TOTAL DEMATIACEOUS MOULDS:	71	0

SPECIES	NO. OF ISOLATES STORED	NO. (%) OF ISOLATES NOT RECOVERED
Dimorphic moulds:		
Blastomyces dermatitidis	5	0
Histoplasma capsulatum	5	0
Penicillium marneffei	17	0
TOTAL DIMORPHIC MOULDS:	**27**	**0**
Dermatophytes:		
Epidermophyton floccosum	4	2
Microsporum canis	12	3
Microsporum gypseum	6	1
Trichophyton mentagrophytes	26	2
Trichophyton rubrum	13	7
TOTAL DERMATOPHYTES:	**61**	15
TOTAL MOULDS:	391	15

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Efficiency of Microbank® for the Conservation of Microorganisms Relevant to Veterinary Medicine

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In this paper the conservation of different bacteria, yeasts and moulds from diagnostic material was examined with a commercial deep freeze system. 137 bacteria isolated from the stomachs of dogs, cats and pigs and from duodenal juice of cats and 7 isolates of yeasts and fungi from diagnostic material were conserved with the deep freeze system Microbank®. Furthermore 62 Helicobacter pylori isolates and 1 Helicobacter felis isolate were conserved with this system. After a storage period of 24 hours up to 20 months the isolates were recultured. 96% of the conserved microorganisms could be grown. It was not possible to cultivate 1 fungus isolate (Fusarium sp.). 2 Helicobacter pylori isolates, spiral bacteria from the stomach of a pig and a Clostridium tyrobutyricum isolate were only recultured after 24 hours, 1 isolate of Moraxella sp. after 2 months and another 4 anaerobe isolates after 5 months. The Microbank® system proved to be suitable for conservation and was also efficient for the conservation of microorganisms, which could not easily be cultivated.

Use of Commercially Available Cryogenic Vials for Long-Term Preservation of Dermatophyte Fungi

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The use of commercially available cryogenic vials (Microbank® vials) stored at -70°C for the storage and preservation of dermatophyte fungi was investigated. None of the 200 strains of dermatophytes examined, representing 21 species, showed a loss of viability after they had been stored for periods ranging from 1 week to 2 years at -70°C. All strains showed typical colonial and microscopic morphologies following revival.

Long-term storage of fungal isolates is critical for preservation of the germplasm and maintenance of stock cultures with minimal effort over long periods. Conservation of morphological, physiological, genetic, and metabolic stability is crucial for many purposes and is vital for isolates used as medical reference strains, for chemotaxonomic studies, or in the commercial production of biochemicals. In a comparative study of the effects of five different storage methods, cryopreservation was the method that best provided for the stability of secondary metabolite production and is now considered the best method available for the long-term storage of microbial cultures. Dermatophytic fungi can present problems for storage, as the cultures often become pleomorphic, with various levels of sporulation or mycelial growth. McGinnis et al. used sterile distilled water to store hyphal and spore suspensions of

147 different species of fungi at 25°C for periods of 12 to 60 months. These included more than 25 species of dermatophytes. The degree of sporulation and the quality of the inoculum appeared to be critical factors; and when the inocula were “adequate” in size, even some poorly sporulating species such as Trichophyton violaceum, Trichophyton schoenleinii, and Microsporum ferrugineum survived storage well. The long-term storage of a wide range of fungal species, including dermatophyte isolates, in commercially prepared cryogenic freezer beads (Microbank®) at -70°C or in liquid nitrogen has been tested. Although most fungi were preserved well by this method, dermatophytic fungi did not show good recovery rates. For example, more than 50% of isolates of Trichophyton rubrum were not recovered. Consequently, this method was not recommended for use for the long-term storage of dermatophytic fungi. In contrast, we have found contradictory results and we report on a simple and successful technique for the long-term storage of dermatophytes.

Fresh isolates were collected from clinical specimens submitted to the Microbiology Department at the William Harvey Hospital, Ashford, United Kingdom, between March 2002 and August 2005. Reference strains were obtained from the National Collection of Pathogenic Fungi, Bristol, United Kingdom. Representative strains of dermatophytes were used to assess a commercially available freezer bead storage kit (Microbank®; Pro-Lab Diagnostics). Each 2-ml tube contains approximately 50 plastic beads (diameter, 3 mm) with a hole through the centre (this hole retains approximately 1 μl of suspension), which allows repeated recovery of an isolate before the preparation of a new stored culture is needed. This is in contrast to traditional long-term storage methods, in which the isolates are stored in multiple single-use vials, and has the added advantage of taking up less space.

Mycelium and conidia were harvested from 7-day-old cultures incubated at 27°C on Sabouraud dextrose agar (SDA; Oxoid Ltd., United Kingdom) by using a sterile scalpel and inoculated into a freezer bead tube containing a suspension medium prepared according to the manufacturer's instructions to give a density approximately equal to or greater than that of a McFarland no. 4 standard. The suspension was shaken vigorously to evenly distribute the fungus and was left to stand for 5 min. Excess fluid was removed with a Pasteur pipette. Before the tubes were frozen and stored at -70°C, a single bead was removed with sterile forceps and was placed on a fresh SDA plate, and the resulting drop of fungal suspension was spread by using a 10-μl loop to obtain single colonies and to check for viability and purity. These plates were incubated at 27°C for 7 days to assess the amount of inoculum present on a single bead. At various time intervals over 24 months, the tubes were removed from the freezer and a bead was removed from the frozen clump, plated, and incubated as described above. The tubes were immediately returned to the -70°C freezer before the contents had thawed. The number of colonies recovered, their growth rate, and the macroscopic and microscopic morphologies of the isolates were noted.

A detailed time course study was conducted with four isolates (Trichophyton interdigitale WHH1268, Trichophyton mentagrophytes WHH692, Trichophyton rubrum WHH3229, and Epidermophyton floccosum WHH1471). Single beads were removed at 0, 1, 2, 3, 4, 5, 6, 7, 8, 12, 16, 20, 24, 32, and 36 weeks and were cultured as described above. Epidermophyton floccosum was included, as it is known to die rapidly if it is kept at 4°C. In all cases, at all time intervals, successful reestablishment of the cultures ensued. At least 1,000 CFU were typically recovered from each bead. The growth rates and the hyphal densities were comparable to those of an initial control culture before it was frozen. Colonial and microscopic morphologies remained true to type throughout.

In addition to this time course study, a second trial was conducted with 58 stored isolates representing 15 species of dermatophytes. The isolates were selected to give a range of species but also to sample a range of isolates within some of these species (e.g., Arthroderma benhamiae, Trichophyton interdigitale, and Trichophyton tonsurans). For this trial, isolates stored for different time periods over the previous 24 months were recultured in triplicate to assess the uniformity of the distribution of viable organisms in frozen tubes. Three freezer beads were taken from each tube of preserved isolates and cultured as described above. In all cases, successful reestablishment from all three replicates occurred for all isolates tested. No adverse effects on morphology or growth rates compared to those of cultures not subjected to cryopreservation were noted.

Following these initial trials, this method of preservation was adopted for the storage of all stock strains in the laboratory. To date, all cultures kept in this manner have been successfully

revived as required at times ranging from 1 week to 2 years, and these cultures represent 200 isolates of 21 species of dermatophytes (Table 1). Espinel-Ingroff et al. have discussed the advantages of using the Microbank® freezer bead system in terms of its availability and ease of use. Their results suggested that dermatophytes would not be well preserved by use of this method. Our results for a wider range of isolates suggest that the limited numbers of dermatophytes that they tested were not representative or that a different preservation technique might have given better recovery rates. For example, half the specimens prepared by Espinel-Ingroff et al. were preserved in liquid nitrogen and kept for up to 8 years, whereas all our specimens were kept at -70°C and tested within 2 years. The recovery of all the isolates used in our study was successful, with no apparent effect on culture phenotype. All four specimens of T. rubrum were recovered in our study, including the isolate used in the detailed time course study. This is in contrast to the 54% recovery rate of T. rubrum isolates in the earlier study. As a consequence, we can now recommend this method of preservation of dermatophytes for clinical laboratories worldwide.

Table 1. Isolates recovered after preservation for 1 week to 24 months

NAME	ISOLATE CODE	NO. OF STRAINS	TIME OF STORAGE (M)
Arthroderma benhamiae	NCPF®410	1	24
A. benhamiae	NCPF®456	1	3
A. benhamiae	NCPF®460	1	0.25
A. benhamiae	Clinical isolates	13	2-10
Arthroderma simii	NCPF®494	1	24
A. simii	NCPF®471	1	1
Arthroderma vanbreuseghemii	NCPF®452	1	24
A. vanbreuseghemii	NCPF®750	1	24
A. vanbreuseghemii	NCPF®749	1	24
Epidermophyton floccosum [b]	Clinical isolate WHH1471	1	24
E. floccosum	Clinical isolates	7	8-24
Microsporum audouinii	Clinical isolates	4	1
Microsporum canis	Clinical isolates	7	9-24
Microsporum gypseum-Microsporum fulvum	Clinical isolates	5	1-9
Microsporum persicolor	NCPF®502	1	24

NAME	ISOLATE CODE	NO. OF STRAINS	TIME OF STORAGE (M)
M. persicolor	Clinical isolates	4	1-9
Trichophyton ajelloi	NCPF®364	1	12
Trichophyton equinum	NCPF®526	1	24
T. equinum	NCPF®565	1	24
T. equinum	NCPF®673	1	18
T. equinum var. autotrophicum	NCPF®488	1	24
T. equinum	Clinical isolate WHH2660	1	24
Trichophyton erinacei	NCPF®652	1	24
T. erinacei	Clinical isolates	10	1-10
Trichophyton interdigitale	NCPF®780	1	24
T. interdigitale	Clinical isolate WHH692	1	24
T. interdigitale	Clinical isolates	55	1-24
Trichophyton mentagrophytes	NCPF®224	1	24
T. mentagrophytes	Clinical isolate WHH1268	1	24
T. mentagrophytes	Clinical isolates	27	1-24
Trichophyton quinckeanum	NCPF®310	1	24
T. quinckeanum	NCPF®341	1	24
T. quinckeanum	Clinical isolate	1	0.25
Trichophyton rubrum	NCPF®113	1	24
T. rubrum	Clinical isolate WHH3229	1	24
T. rubrum	Clinical isolates	2	1-24
Trichophyton schoenleinii	NCPF®691	1	12
T. schoenleinii	Clinical isolate Bristol ST5	1	1

NAME	ISOLATE CODE	NO. OF STRAINS	TIME OF STORAGE (M)
Trichophyton soudanense	Clinical isolates	4	2-14
Trichophyton tonsurans	NCPF®690	1	24
T. tonsurans	Clinical isolates	25	1-22
Trichophyton verrucosum	Clinical isolates	2	1-22
Trichophyton violaceum	Clinical isolates	5	7-22
TOTAL		200	

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Two-Year Study Evaluating the Potential Loss of Methicillin Resistance in an MSRA Culture Collection

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A reported loss of mecA prompted us to monitor 360 cryostocked methicillin-resistant Staphylococcus aureus strains for stability. Concurrently, 14 well-characterised strains were stored in a Microbank preservation system and subjected to multiple freeze-thaw events. There were no significant declines in the methicillin-resistant populations with either method over a two-year period.

Strain stability in a research culture collection is critical for valid and consistent experimental results. The University of South Florida Center for Biological Defense (CBD) stocks well-characterised methicillin-resistant Staphylococcus aureus (MRSA) strains for use in validating molecular typing methods such as multilocus sequence typing, staphylococcal protein A typing, multilocus variable-number tandem repeat analysis, pulsed-field gel electrophoresis (PFGE), and staphylococcal cassette chromosome mec (SCCmec) typing. The CBD collection largely consists of community-acquired strains received from hospitals and clinical laboratories in Florida and Washington State as well as strains obtained from the Network on Antimicrobial Resistance in Staphylococcus aureus (NARSA) and from H. de Lencastre (13). Based on the known association of pulsed-field type with SCCmec type (15), the majority of the strains typed in our collection were USA300 (159 presumed SCCmec type IV), followed by USA100 (103 presumed SCCmec type II), and 84 sporadic pulsed-field types were SCCmec types II, IIA, III, IIIA, and IV and undetermined. The integrity and stability of mecA and SCCmec within our MRSA strains are critical to CBD research.

In 2005, van Griethuysen et al. reported the loss of mecA in an MRSA strain collection after two years of cryostorage at -80°C in a Microbank bead-based preservation system (MBPS) (Pro-Lab Diagnostics, Austin, TX) (16). Prior to storage in the MBPS, the strains were previously held as long-term stocks at room temperature (RT). Two years later, approximately 14% of the collection stored in the MBPS no longer carried mecA.

This report prompted an evaluation of the stability of 360 MRSA strains from the CBD collection. After confirmation of methicillin resistance and phenotypic characterisation from a single, representative colony, each MRSA strain was cryostocked using a standardised cryostorage protocol as follows. The progenitor colony was inoculated to 100 ml of tryptic soy broth (TSB) and incubated at 35°C for approximately 15 h in a shaking water bath at 125 rpm. Thirty millilitres of overnight culture was pelleted at 3,571 × g at RT for 10 min, resuspended in 25 ml of TSB with 10% glycerol, and aliquoted as 1 ml into cryovials. After 30 min of equilibration at RT with the cryoprotectant, cryovials were placed in Nunc controlled freezing units (Nalgene, Rochester, NY) following the manufacturer's instructions and held at -85°C for 75 min to foster cooling of approximately -1°C/min to near -30°C. Immediately thereafter, cryovials were removed from the units and stored at -85°C.

One week after cryostorage, during which most cell death occurs (1), a single cryovial of each CBD MRSA strain was quick-thawed at 35°C and plate counts were obtained from a 1:10 dilution series to review viability and

purity. To confirm that methicillin resistance had been maintained in the population after cryopreservation, disc diffusion susceptibility (DDS) was performed using 30-µg cefoxitin and 1-µg oxacillin antibiotic discs (Oxoid, Hants, United Kingdom) on Mueller Hinton (MH) plates. Zones of inhibition were measured after incubation at 35°C for 24 h, followed by incubation at 30°C for another 24 h. The Clinical and Laboratory Standards Institute (CLSI) standards for these antibiotics were used to interpret the resistance of the strains (3, 4). Control strains for this method and all others used in this study were a methicillin-susceptible Staphylococcus aureus strain, ATCC® 25923 (American Type Culture Collection, Manassas, VA) and a confirmed SCCmec type IV MRSA strain, CBD 804 (HDE288). CBD strains were also tested for the presumptive presence of PBP2', the mecA gene product, from cultures grown at 35°C for 24 h on blood agar following the manufacturers' instructions (Oxoid, Hants, United Kingdom; Denka Seiken, Ltd., Derbyshire, United Kingdom).

At one- and two-year intervals, plate counts were performed on the 360 CBD MRSA strains to ensure that a robust heteroresistant population of MRSA had persisted after prolonged cryostorage at -85°C. CBD MRSA stocks were quick-thawed, and a 1:10 dilution series was spread in 100-µl volumes on tryptic soy agar (TSA) and MH agar containing 4% NaCl and 6 µg/ml oxacillin (MH-OXA) (Remel, Lenexa, KS). By comparing the CFU/ml of the entire population on TSA to that of the MRSA subpopulation on MH-OXA at each yearly time point, we were able to track the stability of the heteroresistant population. All the strains tested produced methicillin-resistant colony counts on MH-OXA equal to or within a log of the heteroresistant CFU/ml on TSA at one- and two-year time points. The means of the counts from year 1 on TSA and MH-OXA were $1.66 \times 109 \pm 4.28 \times 107$ and $1.26 \times 109 \pm 3.14 \times 107$, respectively, and in year 2, $1.89 \times 109 \pm 5.98 \times 107$ and $1.46 \times 109 \pm 3.87 \times 107$ (data not shown). Thus, none of the 360 cryostocked MRSA strains tested from the CBD culture collection demonstrated a significant decline in stability of their heteroresistant populations of MRSA after the two-year review (nonparametric Kruskal-Wallis test; $P = 0.7175$) (Table 1).

During this evaluation of our CBD MRSA collection stability, we also sought to recreate the loss of mecA reported by van Griethuysen et al. for strains stored in the MBPS (16). We subjected six previously described (13) MRSA strains comprised SCCmec types I, IA, II, III, IIIA, and IV and eight PFGE types (USA100 through to -800) obtained from NARSA to two years of cryostorage in the MBPS with periodic freeze-thaw events (Table 1). In contrast to our CBD cryostorage protocol, the MBPS technical instructions directed the inoculation of a cell suspension of young growth for each strain into a cryostorage vial containing 25 porous beads and a proprietary cryoprotectant. The vials were inverted five times and then immediately well aspirated of most liquid. This action gave the cells that had adhered to the beads very little equilibration with the cryoprotectant. The vials were frozen in an uncontrolled manner to -85°C; the manufacturer recommends storage at -70°C.

To monitor the 14 strains for a stable methicillin-resistant subpopulation, a single MBPS bead for each strain was tested at intervals spanning 1 to 6 weeks during the two-year period. At each time point, the MBPS vials were removed from -85°C and held on ice for approximately 10 to 15 min while a bead for each strain was aseptically transferred to 1 ml TSB with 10% glycerol at RT. The MBPS vials were immediately returned to -85°C. Following 5 s of vigorous vortexing of each sacrificed bead per strain, plate counts comparing the heteroresistant population on TSA to the MRSA subpopulation on MH-OXA were performed as described above. The vials containing individual beads suspended in TSB with glycerol were placed directly in -85°C to freeze and were saved for future testing. Within 10 min on ice, the MBPS vials transitioned from approximately -85°C to near 0°C and rose to 5°C during handling to remove a bead, as determined by inserting a temperature probe among the beads. This procedure allowed us to replicate freeze-thaw events.

During cryostorage in the MBPS for 104

weeks, the 14 MRSA strains were subjected to 24 such freeze-thaw events. Thirteen strains did not have a significant decline in the methicillin-resistant population from initial storage in the MBPS to final evaluation at 104 weeks (nonparametric Kruskal-Wallis test; P = 0.1528) (Table 1). The means of the counts from week 0 on TSA and MH-OXA were 5.17 × 106 ± 1.59 × 106 and 2.62 × 106 ± 6.39 × 105, respectively, and in week 104, 8.10 × 106 ± 2.56 × 106 and 4.73 × 106 ± 1.23 × 106 (Table 1). Two strains struggled to tolerate a relatively high concentration of oxacillin from the beginning of the study. CBD 801 (N315), a SCCmec type II strain, repeatedly failed to grow on MH-OXA, and therefore, the methicillin-resistant cells in its population were not enumerated at each time point. CBD 804 (HDE288), an SCCmec type IV strain, also had difficulty thriving on MH-OXA, as it required extended incubation time to produce countable colonies. DDS and PBP2' latex agglutination testing performed at each freeze-thaw event on both strains demonstrated that they were resistant to 1 µg oxacillin and that the mecA gene product was presumptively present.

To investigate whether a decrease in resistance to methicillin occurred among the 14 strains over two years, we examined the stored beads from the first freeze-thaw event, which were collected 1 week after initial storage in the MBPS, and from week 104, in which the beads had undergone the stress of 24 freeze-thaw events. Susceptibility tests using the microbroth dilution method were performed with a Sensititre system (TREK Diagnostic Systems, Cleveland, OH) following the manufacturer's instructions and with CLSI standards (3). A methicillin-susceptible S. aureus ATCC® 29213 strain was the control in all MIC testing. The microtitre plates containing ampicillin, penicillin, oxacillin, and vancomycin were incubated at 35°C for 24 h. The susceptibility tests gave identical or near-identical MICs (a <2-fold dilution difference) on all the antimicrobials for the sacrificed beads stored at week 1 and week 104 for all 14 cryostocked strains in the MBPS.

The presence of mecA was also tested in sacrificed beads stored at week 1 and week 104. Genomic DNA was extracted from overnight bacterial growth in TSB using a MagNa Pure LC instrument (Roche Diagnostics Corp., Indianapolis, IN) with the supplied DNA isolation kit III following the manufacturer's instructions. PCR for mecA was performed as described previously (13). The results confirmed that the mecA gene was present in all 14 strains from their initial storage in the MBPS to the endpoint of our MBPS evaluation.

There are various insults to cells that occur during a freeze-thaw event (11, 12) that may explain the previously reported loss of mecA-positive cells (16). However, a distinction should also be made between the deletion or disruption of mecA and the overgrowth and eventual replacement of MRSA by a coexisting mecA-negative population. MRSA strains are heterogeneous, consisting of two subpopulations wherein the resistant population grows slower than the more-robust susceptible population (2). Stressful environments such as storage at RT and prolonged freezing with repeated freeze-thaw events can play a significant role in evolving lineages that are better able to survive these stresses (14). Therefore, the CBD protocol began with selection and enrichment of a predominantly MRSA population prior to preservation. Confirmation of a robust MRSA population was also done shortly after cryostorage in TSB with 10% glycerol.

The cryostocking process itself could have decreased the mecA-positive subpopulation, since it is known to inflict detrimental changes to bacterial cells, such as ice crystal formation, dehydration, decreased and/or increased activity of enzymes, accumulation of metabolites, increased molecular contacts, disruption of weak hydrogen bonds, distortion of molecules, and breakdown of the cell membrane (12). The CBD cryostocking protocol incorporates osmotic equilibration with a 10% glycerol cryoprotectant. Equilibration with a cryoprotectant minimises damage during freezing by penetrating and stabilising the cell membrane and delaying freezing. While we allow 30 min or more for cells to equilibrate with the glycerol cryoprotectant, the MBPS protocol allows <1 min for cells to equilibrate with the

proprietary cryoprotectant. It is possible that the removal of the cryoprotectant leaves the mecA-positive cells in the MBPS more vulnerable to injury during freeze-thaw events. Also, the MBPS technical instructions do not specify how to handle the vials while retrieving beads. The injurious complexities of a freeze-thaw event stem largely from uncontrolled rates of cooling and warming during which ice in the cells (re)crystallises (8, 11). The MBPS vials in our experiment were held on ice during bead retrieval, in keeping with a common practice that results in relatively rapid thawing and reduced cell injury (8, 11). Our standardised protocol includes a programme of controlled, slow freezing to minimise intracellular ice formation, followed by uncontrolled, rapid freezing to forestall further dehydration.

We did not detect a significant decline in 13 MRSA populations in the MBPS after 24 freeze-thaw cycles in two years; the MRSA population for CBD 801 (N315) could not be enumerated on MH-OXA after initial evaluation, but methicillin resistance was confirmed at each time point. However, it is possible that low-temperature storage or multiple uncontrolled freeze-thaw events give "freeze-hardy" methicillin-susceptible cells in the heterogeneous population a "cryopreservation selection" advantage over the methicillin-resistant subpopulation during thawing and reculturing (11). To avoid these potential problems, we stock multiple single-use cryovials for every strain in our collection and perform quality control checks with each use.

A previous study reported that genetic background affects the stability of mecA in MRSA (10). The distribution of SCCmec contributes to the efficiency and stability of PBP2' expression, which propagates methicillin resistance. The mecA gene is well maintained on an engineered plasmid in methicillin-susceptible Staphylococcus aureus strains from major clonal complexes.

Table 1: Plate counts of heteroresistant and methicillin-resistant populations

CBD NO.		DESCRIPTION	EVALUATION OF CBD CRYOSTOCKS			
			YEAR 1		YEAR 2	
			TSA	MH-OXA	TSA	MH-OXA
799[c]	COL	SCCmec type I	2.0×10^9	2.1×10^9	2.4×10^9	2.0×10^9
800	PER34	SCCmec type IA	1.5×10^9	1.3×10^9	1.7×10^9	1.4×10^9
801	N315	SCCmec type II	1.3×10^9	NG[f]	1.3×10^9	NG
802	ANS46	SCCmec type III	1.9×10^9	1.8×10^9	1.6×10^9	1.1×10^9
803	HU25	SCCmec type IIIA	1.6×10^9	1.8×10^9	2.0×10^9	2.1×10^9
804	HDE288	SCCmec type IV	6.4×10^8	3.0×10^8	1.4×10^5	7.8×10^8
1064[d]	NRS 382	USA100 (II)[e]	2.4×10^9	1.7×10^9	6.1×10^9	8.5×10^8
1065	NRS 383	USA200 (II)	1.3×10^9	1.3×10^9	2.6×10^9	2.2×10^9
1066	NRS 384	USA300 (IVa)	1.8×10^9	3.5×10^8	1.2×10^9	1.1×10^9
1067	NRS 123	USA400 (IVa)	4.3×10^9	1.1×10^9	2.2×10^9	2.0×10^9

CBD NO.		DESCRIPTION	EVALUATION OF CBD CRYOSTOCKS [a]			
			YEAR 1		YEAR 2	
			TSA	MH-OXA	TSA	MH-OXA
1068	NRS 385	USA500 (IV)	1.4×10^9	9.9×10^8	1.3×10^9	1.2×10^9
1069	NRS 22	USA600 (II)	5.9×10^8	2.8×10^8	5.7×10^8	6.9×10^8
1070	NRS 386	USA700 (IVa)	1.3×10^9	9.3×10^8	2.6×10^9	3.0×10^9
1071	NRS 387	USA800 (IV)	3.3×10^9	1.1×10^9	1.2×10^9	1.0×10^9
799c	COL	SCCmec type I	1.8×10^6	1.6×10^5	7.8×10^6	6.0×10^6
800	PER34	SCCmec type IA	3.4×10^6	2.1×10^6	4.6×10^6	4.5×10^6
801	N315	SCCmec type II	5.6×10^6	NG	6.0×10^6	NG
802	ANS46	SCCmec type III	1.8×10^7	1.5×10^7	1.5×10^6	1.7×10^7
803	HU25	SCCmec type IIIA	7.6×10^6	8.3×10^6	9.6×10^6	9.6×10^6
804	HDE288	SCCmec type IV	8.4×10^6	5.2×10^6	7.9×10^6	1.4×10^5
1064d	NRS 382	USA100 (II)e	2.5×10^6	2.1×10^6	1.7×10^6	2.0×10^7
1065	NRS 383	USA200 (II)	1.0×10^5	7.6×10^6	1.9×10^6	2.1×10^6
1066	NRS 384	USA300 (IVa)	1.9×10^6	4.3×10^6	1.6×10^7	1.4×10^7
1067	NRS 123	USA400 (IVa)	5.8×10^6	1.0×10^6	1.2×10^7	1.2×10^7
1068	NRS 385	USA500 (IV)	4.6×10^6	4.5×10^6	9.6×10^6	8.1×10^6
1069	NRS 22	USA600 (II)	1.0×10^6	1.4×10^6	3.2×10^6	1.9×10^6
1070	NRS 386	USA700 (IVa)	1.0×10^7	5.4×10^6	1.8×10^7	1.3×10^7
1071	NRS 387	USA800 (IV)	1.5×10^6	9.6×10^5	1.6×10^7	1.7×10^6

a Representative data in CFU/ml for 14 of 360 MRSA strains cryostocked with a standardized protocol in the CBD collection.
b Representative data in CFU/ml from 14 well-defined MRSA strains stored in the MBPS.
c CBD 799 to 804 were derived from strains previously described (10).
d CBD 1064 to 1071 were derived from NARSA strains.
e Pulsed-field type (SCCmec type). SCCmec type information obtained from NARSA.
f NG, no growth. Strain confirmed for methicillin resistance by DDS and PBP2' testing.

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An Internal Quality Assurance Scheme for Clinical Bacteriology using Microbank®

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The development of an internal quality control scheme in clinical bacteriology has been hampered by a lack of suitable cultures. However, work undertaken using Microbank® beads may provide a solution to this problem.

Internal Quality Assessment in Clinical Bacteriology

One definition of quality involves meeting the predetermined requirements of users of a product or service. An effective quality management system (QMS) determines the needs and expectations of users and evaluates the processes, responsibilities and resources required to meet quality objectives. In the laboratory, quality control (QC) procedures should be used in conjunction with external and internal quality assessment (QA), audit and equipment monitoring as an integral part of the QMS. Quality control permits the day-to-day monitoring of assay, operator and equipment performance. It should detect both random and systematic errors.

Criteria for Quality Control Material

In general, QC material should be independent of kit controls, be stable over a long period of time, be of sufficient volume to monitor within and between kit and reagent and batches, and give results within a clinically significant range (for bacteriological cultures, this means target organisms).

Unfortunately, availability of suitable QC material presents a problem in developing a suitable internal QA scheme in clinical bacteriology. One scheme established for QA in clinical bacteriology involves the submission of anonymised original specimen for analysis(1); however, problems associated with this type of QC material include:

- Failure to meet at least two of the criteria mentioned above.
- Repeat inoculation of a swab on a second set of plates may present a different picture (depending on the number of organisms originally present).
- A high percentage of bacteriology samples are negative and this does not challenge the ability to isolate 'target' organisms.

Although this type of scheme assesses reproducibility, it does not detect systematic errors (because you don't know what you might be missing).

Criteria for Quality Control Material

An alternative scheme using simulated specimens preserved on Microbank® beads (Pro-Lab Diagnostics) was evaluated. Microbank® offers a ready-to-use design to simplify the storage and

retrieval of bacterial and fungal cultures. Comprising a special cryovial system that incorporates treated beads and a cryopreservative solution, Microbank® provides a more reliable means of maintaining cultures than is possible with repetitive subculture, which can result in contaminated cultures, lost organisms or changed characteristics. The special formulated preservative ensures longer survival of fastidious organisms and higher quantitative recoveries. This makes the Microbank® system ideal for QC applications where organism integrity, consistency and quality are of paramount importance.

Each 2 ml Microbank® vial contains approximately 25 beads, providing the facility for repeat culture of the original organism using a simple procedure. Extensive, proven performance reference data, updated for 2005, is available on request from Pro-Lab Diagnostics.

To demonstrate the utility of the Microbank® system, seven simulated specimens were prepared. Briefly, using freshly isolated colonies, a suspension of the target organism was prepared (equivalent to a McFarland 2 standard) in a Microbank® vial. To simulate a clinical specimen, colonies of typical mixed normal flora were added to make the final suspension equivalent to a McFarland 5 standard (Table 1). The vial was mixed thoroughly and the contents were decanted into a Petri dish. Using sterile forceps, each bead was placed in an individual cryotube, which was then labelled and stored frozen at -80°C.

Table 1: Simulated Specimens

NO	SPECIMEN TYPE	TARGET ORGANISM(S)	OTHER ORGANISMS
1	Throat Swab	Group A streptococcus	Mixed oral flora
2	Throat Swab	Corynebacterium diphtheriae	Mixed oral flora
3	Sputum	Haemophilus influenzae	Mixed oral flora
4	Sputum	Streptococcus pneumoniae	Mixed oral flora
5	Wound Swab (burn)	Staphylococcus aureus Pseudomonas aeruginosa	Coagulase-negative staphylococcus
6	Faeces	Group D salmonella	Escherichia coli Proteus mirabilis
7	Faeces	Escherichia O157	Escherichia coli (sorbitol-positive)

Culture Results

Over a six-month period (June to December 2003) all target organisms were isolated and correctly identified from six out of the seven simulated specimens. On two occasions, isolation of Haemophilus influenzae from specimen 3 failed.

Overview

Once prepared, simulated samples are simple to set up, record and score, and are inexpensive to prepare. In addition, they satisfy all the criteria for a QC material. Drawbacks include the fact that simulated samples are not 'real' specimens, and that some organisms may survive better than others at -80°C after 'pooling'. Fastidious organisms such as Neisseria gonorrhoeae, Campylobacter spp. and anaerobic organisms have yet to be tested using the Microbank® system.

It may be argued that staff would soon get to know which organisms are present in the samples, but this argument also can be applied to most QC material used across pathology and is not relevant unless a 'blame culture' exists in the organisation. Used properly, however, a successful internal QC scheme will increase confidence in results and, in conjunction with external QA and audit, identify problems and assess the effectiveness of remedial measures.

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Germany Data - Bestbion DX

Following many years of successful application of the Microbank® system for the long-term storage and retrieval of bacterial and fungal cultures in many German microbiology laboratories and institutes, a request was made for inclusion of this data in the Microbank® Worldwide Performance Portfolio. Our exclusive distributor in Germany for the Microbank® system (Bestbion dx. www.bestbion.com) conducted an extensive market survey inviting laboratories to submit data showing the successful use of the Microbank®. The following laboratories kindly contributed to the summary that follows below. We are grateful to all of our German customers and Bestbion dx for the continued support and confidence in the Microbank® product.

Participating Laboratories:

Sanitätsakademie d. Bundeswehr München, Institut für Mikrobiologie
Diakonissenkrankenhaus Flensburg, Zentrallabor
MVZ Labor Schweinfurt, Bakteriologie
Hamburger Wasserwerke, Wasserlabor-Mikrobiologie
Eberhard-Karls-Universität Tübingen, Institut für Tropenmedizin
Klinikum Ansbach, Bakteriologie
Klinikum Bamberg, Institut für Labormedizin
Klinikum Kulmbach, Zentrallabor
Klinikum St. Marien Amberg, Zentrallabor
LaboKlin GmbH & Co. KG, Labor für klinische Diagnostik
Labor Bamberg
Labor L+S AG
LADR GmbH MVZ Dr. Kramer & Kollegen
Max Rubner-Institut Kulmbach
Labor an der Salzbrücke MVZ GmbH
Medizinisches Versorgungszentrum Dr. Stein u. Kollegen
Medizinisches Versorgungszentrum Dres. Cornely, Riebe & Berndt
Tierärztliche Gemeinschaftspraxis Dres. Windhaus & Hemme
Universitätsklinikum Magdeburg, Institut für Medizinische Mikrobiologie und Krankenhaushygiene
Universitätsklinikum Heidelberg, Medizinische Mikrobiologie und Hygiene
Universitätsklinikum Ulm, Institut für Medizinische Mikrobiologie und Hygiene
Universitätsklinikum Würzburg, Klinik u. Poliklinik f. Dermatologie, Mykologielabor
Westpfalz-Klinikum Kaiserslautern, Institut f. Laboratoriums- und Transfusionsmedizin

Results: Storage Temp -20°C

ORGANISM/SPECIES	≤12 M	≤24 M	≤36 M	≤48 M	≤60 M	> 5 Y
Bacteroides spp.	2		1			1
Bordetella spp.					1	
Campylobacter jejuni	1		1			1
Candida spp.		1	1			1

ORANISM/SPECIES	≤12 M	≤24 M	≤36 M	≤48 M	≤60 M	> 5 Y
Clostridium spp.	2		2			1
EHEC		1				0
Escherichia coli	1		2			1
Haemophilus spp.	1		1			1
Klebsiella pneumoniae	1					0
Listeria monocytogenes				1		0
Myroides odoratus						1
Neisseria gonorrhoeae	1		1			0
Ochrobactrum anthropi					1	0
ORT			1			0
Pasteurella spp.					1	0
Pasteurella multocida						1
Pseudomonas aeruginosa	1					0
Riemerella spp.		1				0
Rhodococcus equi						1
Staph. aureus / MRSA			1	1		3
Streptococcus Group B		1	1			1
Streptococcus pneumoniae	1		1			1
Taylorella equigenitalis				1		0
Yersinia enterocolitica		1			1	0

Results: Storage Temp -40°C

ORGANISM/SPECIES	≤12 M	≤24 M	≤36 M	≤48 M	≤60 M	> 5 Y
Bacteroides spp.	1	1				1
Campylobacter jejuni	2					1
Candida spp.	1	1			1	1
Clostridium spp.	2	1				1
Escherichia coli		1	1		1	1
Haemophilus spp.	3					1
Helicobacter pylori				1		0
Mycobacterium fortuitum						1
Neisseria gonorrhoeae	3					0
Staph. aureus / MRSA		1	1		1	1
Streptococcus Group A						1
Streptococcus Group B	1		1		1	0
Streptococcus pneumoniae	3					1

Results: Storage Temp -70°C

ORGANISM/SPECIES	≤12 M	≤24 M	≤36 M	≤48 M	≤60 M	> 5 Y
Aspergillus spp.	1					0
Bacteroides spp.	1		1			2
Campylobacter jejuni	1			1	1	1
Candida spp.	1	1			1	2
Clostridium spp.	1					2
Enterococcus faecalis	1					1
Escherichia coli	3	1				4
Haemophilus spp.		1		1	1	2
Helicobacter pylori				1		0
Mycobacterium spp.				1		0
Mycobacterium tuberculosis	1			1		0
Neisseria gonorrhoeae		1			1	2
Propionibacterium acnes	1					0
Pseudomonas spp.						1
Pseudomonas aeruginosa	1					0
Pseudomonas fluorescens			1			0
Staph. aureus / MRSA	1	1		1		4
Streptococcus Group B		1			1	3
Streptococcus pneumoniae		1			1	3
Yersinia spp.			1			0

Results: Storage Temp -80°C

ORGANISM/SPECIES	≤12 M	≤24 M	≤36 M	≤48 M	≤60 M	> 5 Y
Aspergillus spp.						1
Bacteroides spp.						1
Blastomycetes						1
Campylobacter jejuni	1					1
Candida spp.						1
Clostridium spp.			1			1
Cryptococcus albidus						1
Dermatophytes						1
Escherichia coli						1
Enterococcus faecalis						1
Haemophilus spp.	1	1				0
Helicobacter pylori	1	1				0
Listeria monocytogenes						1
Moraxella catarrhalis						1
Mycobacterium tuberculosis		1				0
Neisseria gonorrhoeae	2					0
Nocardia spp.						1
Staph. aureus / MRSA				1		1
Streptococcus Group B		1				1
Streptococcus pneumoniae	1					1

Storage, Archiving and Retrieval of Bacteria or Fungi: an overview

Biobanking increases in importance as scientific development and research continues. Here, Mark Reed offers a brief, not-on-the-high-street, insight into what is now a complex science. Biobanking in general terms describes the practice of storing material of a biological origin safely over short or long periods of time. It offers a resource for research, analysis, epidemiological and statistical analysis, and education from large and diverse population sources in numerous scientific applications in many areas and disciplines of science. Samples can be used for individual reference and study or by multiple international groups where access is granted. The term is used in various ways and a simple approach is to define it as a collection of organised samples of human, plant, animal or microbial biological materials accompanied by associated information stored for one or more research purposes.



Biobanks usually incorporate cryogenic storage facilities that vary in size.

Since the 1990s, biobanks have become an important resource in medical research, supporting new approaches such as genomics and personalised medicine. Millions of biological samples, including cells of human, animal or bacterial origin, viruses, serum/plasma or DNA/RNA, are now stored every year around the world for diagnostics and research. In 2002 the expansion and importance of biobanking was recognised by the Medical Research Council (MRC), the Wellcome Trust and the Department of Health, leading to support and comment from the then Prime Minister.

One clear example is for disease research associated with single nucleotide polymorphisms (SNPs) and being able to perform genome-wide association studies using large collections of samples that represent tens or hundreds of thousands of individuals. This can help to identify disease biomarkers, and has led to many important findings in clinical research. Prior to the advent of biobanks, researchers may have struggled to acquire sufficient samples to carry out this important research.

Reference: Storage, Archiving and Retrieval of Bacteria or Fungi: an overview, Pathology in Practice, Mark Reed, August 2018.

Of course, biobanks have also provoked questions on research practices, research ethics, medical ethics and privacy considerations, not to mention important considerations for GDPR. Very careful attention to these points must be given, and to government principles, when setting up a biobank or considering the use of a commercial service or product. As with many areas of scientific research, viewpoints will differ, but it is a fact that numerous biological samples are stored in biobanks from many sources (Table 1). As readers will be aware, in 2009 Clinical Pathology Accreditation (UK) became a wholly owned subsidiary of the United Kingdom Accreditation Service (UKAS), and all laboratories are now in transition to comply with the internationally recognised standard ISO 15189:2012, and the matter of the quality, reliability, security, management and efficiency of biological samples is important for successful accreditation.

Table 1. Examples of biological samples stored in biobanks.

- Biological fluids
- Blood and serum samples
- Tissues
- Organs
- Scrapings
- Spores
- Bacteria
- Viruses
- Parasites
- Cell lines
- Stem cells
- Slide preparations

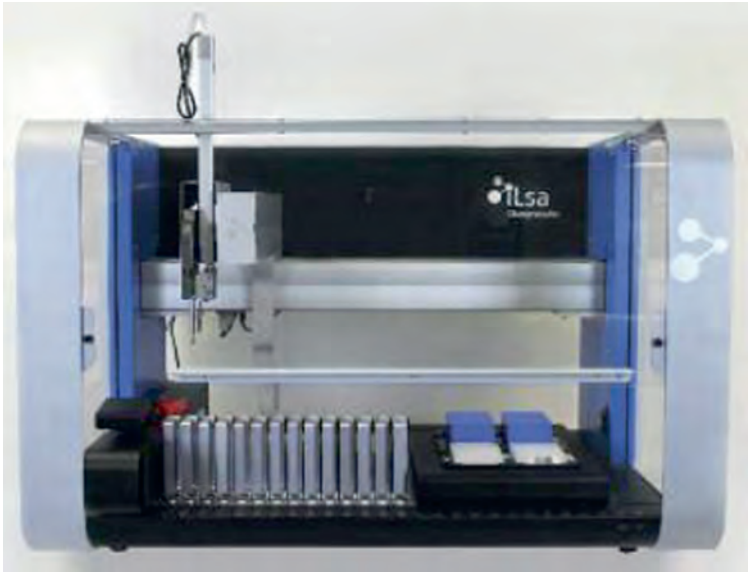
The storage of serum samples represents a significant proportion of the biobanking undertaken internationally. Laboratory-based diagnostic tests using blood and serum samples account for almost 65% of the sample types submitted to pathology departments on a daily basis. Sample numbers can often run into thousands each day, depending on the population area served. Automation in this area of clinical laboratory testing is moving at a tremendous pace and the appropriate biobanking of the samples has to be very carefully considered and planned.

Manual preparation of the samples and storing at -20°C to -70°C can be time-consuming, costly and carry a risk of human error, and there are now a number of automated platforms available to manage this process efficiently. When selecting an appropriate platform, consider improving productivity and ensure the system not only meets current needs but also future needs, allowing for significant increases in sample numbers as the UK laboratory landscape continues to change with amalgamation and centralisation of services

There will be pressures on time and productivity, and the solution chosen must provide a complete and reliable liquid storage management system for biological sample transfer from the primary tubes to secondary tubes and plates, guaranteeing traceability and security. One such example is the new NEO Sample Management System (Ilsa, France. Fig.2.), which is designed to guarantee the traceability of samples, to ensure sample integrity, to improve tracking efficiency and finally to optimise freezer space in order to manage biobank needs effectively.



Diagnostic tests using blood and serum samples account for almost 65% of the sample types submitted to pathology departments.



The NEO system uses integrated 2D Datamatrix GS1-compliant barcodes and the NEO Manager software to guarantee traceability and efficient sample retrieval from the biobank.

The NEO Sample Management System manages the full life-cycle of the sample, from reception of the primary tubes to expiry or final analysis of the resulting aliquots. The refrigerator or freezer storage phase is also fully monitored. The link between the primary sample tube number, the patient number and the well number are all entered electronically, creating an unbreakable link, and only a barcode reader is needed to locate a microtube for a particular patient.

The NEO system uses integrated 2D DataMatrix GS1-compliant barcodes, together with the NEO Manager (software), which guarantees traceability and efficient retrieval from the biobank. NEO Sample Management is applicable for clinical, serum bank, blood bank, cord blood, diagnostics, drug centre, biotechnical, veterinary and safety testing laboratory applications.

As biobanking progresses, public, government and private non-profit organisations are pursuing nationwide and international programmes. One example is the Swedish National Biobank Programme (http://www.biobank.se), a joint national programme on functional genomics. It aims to increase the knowledge and the quality of the Swedish biobanking system, to increase usability and availability of stored samples, and to increase ethical awareness. Another example of a nationwide programme is the United Kingdom Biobank (www.uk.biobank.ac.uk). Such organisations offer a major resource to support a diverse range of research initiatives, which can, in turn, improve the prevention, diagnosis and treatment of disease.

Storage of Living Cells

Consider now the storage of living cells in a biobank. The intention here is not to maintain the integrity of a measurable parameter in a biological sample, but to retain living cells with genotypic and phenotypic characteristics intact over a short or preferably long period of time at -20°C to -70°C. The storage of bacterial and fungal isolates presents its own challenges and is faced with all of those encountered with biological samples, such as security, reliability and compliance.

Established collections of microorganisms permit the characterisation of microbial diversity and microbial evolution, and as microorganisms are essential parts of the biosphere then secure and effective storage is essential. Microbial and fungal culture collections face a huge task: consider that there are potentially over 1.5 million fungal species estimated worldwide, and less than 20% have been described to date.

Culture collections range from localised research projects to laboratory quality control collections, to national and international resource collections. Regardless of the size of microbial and fungal cultures, biobanks are an invaluable tool for accelerated discovery and characterisation of microorganisms and for promoting their beneficial use.

Culture collections must be maintained reliably with minimal deterioration over time, and they must be protected from physical damage, both accidental and intentional, and the registration of each sample should be stored centrally, usually on a computer-based system that can be backed up frequently. Today, there are millions of microbial and fungal cultures stored in culture collections around the world. There are approximately 580 culture collections in 68 countries registered in the World Data Centre for Microorganisms (http://wdcm.nig.ac.jp); set up as a data centre for the World Federation for Culture Collections, it holds in excess of one million microbial strains, of which 44% are fungi, 43% bacteria, 2% viruses, 1% cells, and 10% other agents.

A continuous evaluation at the Anaerobic Reference Centre (Cardiff) has demonstrated that the ***Microbank® can offer a long-term solution.***

Familiar members are the National Collection of Type Cultures, founded in 1920, the longest-established collection of its type anywhere in the world that also serves as a United Nations Educational, Scientific and Cultural Organization Microbial Resource Centre; the Leibniz-Institut DSMZ (Deutsche Sammlung von Mikroorganismen und Zellkulturen) containing over 31,000 cultures represents some 10,000 species and 2000 genera in total, making it one of the largest in the world; and the American Type Culture Collection in the United States. All these organisations offer invaluable service to the science community, providing resources of reference microorganisms and cell lines for research and diagnostic purposes.

Specific conditions are required for the successful long-term storage of microbial and fungal cultures as a biobank. The most widely used systems for storage include cryopreservation and freeze-drying. Cryopreservation means that materials are stored at low (from -20°C to -80°C in freezers) or ultra-low temperatures (-150°C in liquid nitrogen containers); in the latter case, cryopreservation takes place in the liquid or in the vapour phase of nitrogen. Freeze-drying can present practical hurdles and is not ideally suited to smaller, localised culture collections.

Feltham et al. developed a simple method for storage of bacteria at -70°C. Their work revolutionised the way bacteria and fungi could be stored with simple preparations of a glycerol broth using small carrier beads in a vial that could be prepared by individual laboratories. This had a limited scale of application as the need for cultures to be stored for research, quality control and reference applications increased. The advent of International Organization for Standardization (ISO) regulatory requirements for diagnostics manufacturers and laboratories now requires a far more sophisticated product.

Success of Microbank®

Microbank®, manufactured by Pro-Lab Diagnostics, has offered a solution to laboratories for the efficient storage of bacterial cultures for over 28 years. It is an advanced platform that utilises specially etched porous glass beads and a uniquely formulated cryopreservative.

On-going independent laboratory trials have accumulated up to 25 years of successful storage data for both bacterial and fungal cultures, including fastidious bacteria such as Neisseria gonorrhoeae, Campylobacter spp. and anaerobes. More recently, the system has been enhanced with a 2D DataMatrix GS1-compliant barcode to meet higher requirements for traceability and data storage. All 2D barcodes are unique, with a guarantee of no duplicates, and are laser applied for high contrast, and permanently fixed to the tube for guaranteed sample traceability.

Each vial contains approximately 25 sterile coloured beads (single colour) and the unique cryopreservative. The specially treated beads are of a porous nature allowing microorganisms to readily adhere to the bead surface.

After inoculation, the vials are kept at -20°C to -70°C for extended storage. When a fresh culture is required, a single bead is removed from the vial and used directly to inoculate a suitable culture medium. The 2D barcodes on the box and vials provide a freezer-safe, non-erasable label that can be scanned into the laboratory information system (LIS) with ease. This feature not only removes the risk of transcription errors but also has the added advantage of making retrieval from the freezer a simple task. Secure traceability can be enhanced with a customer-specific Microbank® Assets Web Database.

Particular attention must be paid to the use and selection of the storage system when used as a bacterial biobank with fastidious bacteria and in particular anaerobic bacteria. Their primary culture requires specialist culture media and incubation conditions; their storage requires the same dedication.

*Microbank®, manufactured by Pro-Lab Diagnostics, has offered a solution to laboratories for the efficient storage of bacterial cultures for over **28 years**.*

Evaluation in Cardiff

A continuous evaluation at the Anaerobic Reference Centre (Cardiff) has demonstrated that the Microbank® can offer a long-term solution.

Vials were inoculated with clinical isolates of obligatory anaerobic bacteria referred to the Anaerobe Reference Unit (now the National Public Health Service for Wales Anaerobe Reference Laboratory [ARL]) for confirmation of identity, and were frozen at -80°C. A total of 100 isolates were selected to represent the range of species commonly isolated from clinical material (Table 2). One bead from each vial had been removed to demonstrate viability after five years, seven years and 10 years, although beads may have been removed on other occasions

Table 2. Microorganisms for study (n=100) selected to represent the range of species commonly isolated from clinical material.

Actinomyces spp. (2)	*Actinomyces gerencseriae*	*Actinomyces israelii* (2)
Actinomyces naeslundii (2)	*Actinomyces odontolyticus*	*Actinomyces turicensis*
Actinomyces viscosis	*Bacteroides distasonis* (2)	*Bacteroides fragilis* (4)
Bacteroides opvatus (2)	*Bacteroides splanchnicus* (2)	*Bacteroides thetaiotaomicron* (3)
Bacteroides uniformis	*Bacteroides vulgaris*	*Bifidobacterium* spp. (2)
Bifidobacterium animalis group	*Bifidobacterium longum*	*Clostridium bifermentans*
Clostridium butyricum/beijerinckii	*Clostridium cadaveris*	*Clostridium clostridioforme*
Clostridium difficile	*Clostridium glycolicum*	*Clostridium novyi* type A
Clostridium paraputrificum	*Clostridium perfringens* (2)	*Clostridium septicum*
Clostridium sordellii	*Clostridium sporogenes*	*Clostridium tetani*
Clostridium ramosum	*Eggerthella lenta (Eubacterium lentum;* 3)	*Eubacterium aerofaciens* (2)
Peptostreptococcus spp. (3)	*Peptostreptococcus anaerobius* (2)	*Peptostreptococcus asaccharolyticus* (2)
Peptostreptococcus micros (2)	*Peptostreptococcus magnus* (2)	*Peptostreptococcus productus* (2)
Propionibacterium acnes (2)	*Staphylococcus saccharolyticus*	*Streptococcus mutans*

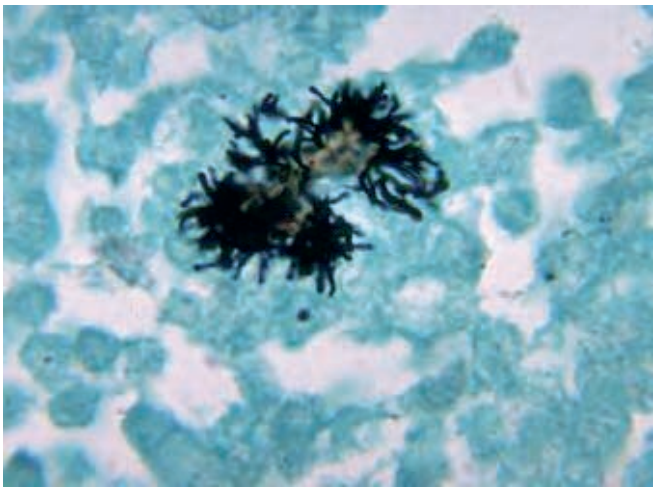
In summary, the evaluation demonstrated that Microbank® vials are easy to use, compact, maintain viability and, therefore, ***are convenient for the long - term storage...***

One bead from each vial was placed aseptically onto Anaerobe Basal Agar (Thermo Fisher, Basingstoke) containing 5% horse blood. Plates were spread for single colonies and were incubated promptly at 37°C in an anaerobic chamber (Ruskinn Concept Plus) for 48 hours. On removal from the chamber, plates were examined for growth of colonies characteristic of the intended isolates. If growth was not apparent at this stage, plates were re-incubated for a further three days.

All 100 strains of anaerobic bacteria of clinical origin were viable after storage in Microbank® vials at -80°C for 10 years. Most strains yielded moderate to heavy growth from a single bead after 48 hours' incubation, but three strains yielded only light growth after five days' incubation. These comprised one of the two strains of Prevotella denticola examined, one of four Fusobacterium nucleatum strains and one strain of Fusobacterium varium.

One hundred anaerobic bacteria representing a wide range of genera isolated from clinical sources remained viable after 10 years' storage in Microbank® vials at -80°C. The same 100 vials were sampled previously for viability after five and seven years' storage. However, 34 of those 100 isolates had changed names since their original identification. These changes reflect advances in taxonomy and identification methods over the decade.

Some species have simply been placed in novel genera and, in some cases, have changed gender in the process (e.g. Eubacterium lentum became Eggerthella lenta). The Gram-positive anaerobic cocci have undergone major taxonomic review, resulting in the removal of most former Peptostreptococcus species to novel genera (e.g. Finegoldia, Anaerococcus, Peptoniphilus). Additionally, several novel species of anaerobic cocci have been described (e.g. Peptoniphilus harei, Peptoniphilus ivorii). Several novel species have been described in other genera (e.g. Prevotella, Porphyromonas, Actinomyces). The identification methods used in laboratories have also been revised to accommodate such taxonomic changes.



Novel molecular methods have been developed for Actinomyces (pictured) and other non-sporing Gram-positive bacilli (Silver stain)

In addition, novel molecular methods have been developed, such as amplified 16S ribosomal DNA restriction analysis (ARDRA) for identification of Bacteroides, Prevotella and Porphyromonas, and for Actinomyces and other non-sporing Gram-positive bacilli. Consequently some strains examined in the Microbank® storage trial have been redesignated as a result of retrospective identification. By chance, two strains previously included in the Microbank® storage trial as Actinomyces species subsequently have been identified as members of these novel species.

In summary, the evaluation demonstrated that Microbank® vials are easy to use, compact, maintain viability and, therefore, are convenient for the long-term storage of anaerobic bacteria. The vials are particularly practical for repeated retrieval of strains as they ensure that the same strain is subcultured on each occasion with minimal effort.

The ARL holds a collection of approximately 20,000 referred isolates dating back to the early 1980s. Isolates received in the past decade have been stored in Microbank® vials. This collection is a valuable resource for retrospective research in fields such as the evaluation of novel identification methods, monitoring of antimicrobial susceptibilities, and development of molecular typing schemes. The author is grateful to Dr Val Hall, Anaerobe Reference Laboratory, for help and support.

Further Reading

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- Seidel KE, Gareis M. Efficiency of Microbank® systems for the conservation of microorganisms relevant to veterinary medicine and others which are not easy to cultivate (in German). Berl Munch Tierarztl Wochenschr 1995; 108 (6): 215-20.
- Williams NJ et al. Microbank®. World Wide Performance Portfolio (www.pro-lab.co.uk/media/custom/upload/File-1455200521.pdf).
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Nottingham University Hospitals NHS Trust Pathogen Bank

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The Nottingham University Hospitals NHS Trust Pathogen Bank initially is an NIHR-funded initiative to collect and store clinically relevant pathogens for their use in three key areas:

- Product development – diagnostic assays, medical devices, antimicrobial testing.
- Basic research - academia and industry
- NHS laboratory requirements - validation and verification, training and development of staff, IQA

This has been achieved by utilising the requirement of clinical microbiology laboratories that provide a diagnostic service to store clinically significant isolates for 12 months. After this storage period these isolates are usually discarded, however we are sorting through these isolates and selecting interesting (i.e. rare strains, highly resistant organisms etc.) for storage in the Pathogen Bank. To give us a varied selection of organisms that might be of interest to multiple organisations we initially stored many common strains as well those with varying antibiograms. We currently have ~3.5K isolates of 94 different bacteria and yeast genera isolated from a number of different diagnostic specimens including but not limited to urine, stool, sputum, blood cultures and pus. All of our strains are stored on Microbank® beads (Pro-Lab Diagnostics) and stored at -70°C for long-term storage.

Once identified as candidates for the Pathogen Bank isolates are cultured from the diagnostic storage using suitable techniques for each organism. The species is then confirmed using the Microflex MS MALDI Smart Analyser with FlexControl software version 3.4 (Bruker Daltonics) with a score of at least 2.0. If the identification provided matches the identification provided in the diagnostic workup to genus level the isolate is included in the Pathogen Bank. Currently the only clinical information we store with the isolates is diagnostic sample type, sample description and antibiotic profile. If organisations require more clinical information ethical approval would have to be considered on a case by case need; however, this is available through our NHS trust and pathology systems.

Quality assurance of strains is maintained using a number of systems, firstly by continual temperature monitoring by Comark. Secondly, each organism stored has two vials, one for use and one as a reserve. These are stored in different freezer sections, each of which have a different power supply to minimise the loss of function if there is a mechanical issue or user error. Cultures are not re-streaked for extended to prevent the introduction of point mutations over time. Lastly, each week a random selection of organisms is tested for viability and given a repeat MALDI-TOF identification. Discrepancies are fully investigated to maintain quality and processes are fully audited as part of the laboratory quality management system.

A repeat MALDI-TOF identification is performed when an isolate is requested to ensure the user receives what they are expecting. Isolates can be sent to users in a number of ways to suit individual needs. If an organisation has a specific requirement for an organism not currently in the Pathogen Bank we can collect isolates prospectively but cannot guarantee how long this process will take.

Services that we offer

- Isolates directly from the Pathogen Bank
- Prospective collection of isolates not in the Pathogen Bank
- Disc diffusion (EUCAST) antibiotic susceptibility testing
- ETEST antibiotic susceptibility testing
- MALDI-TOF identification of isolates provided to us (not for retention in the Pathogen Bank)

The Pathogen Bank has been utilised to generate data to gain CE-IVD accreditation for a number of diagnostic products to date. Strains have also been used in basic research and have led to peer-reviewed publications. Locally, strains have been used in the validation and verification process for the introduction of new diagnostic assays and the training of biomedical scientists and clinical registrars.

For more information contact any of the Pathogen Bank team on:
Pathogenbank@nuh.nhs.uk

List of bacterial and yeast genera available in the Pathogen Bank.

A

- Abiotrophia
- Achromobacter
- Acinetobacter
- Actinomyces
- Actinotignum
- Aerococcus
- Aeromonas
- Aggregatibacter
- Alcaligenes
- Alistipes
- Anaerococcus
- Arcanobacterium
- Atopobium

B

- Bacillus
- Bacteroides
- Bifidobacterium
- Bordetella
- Brevibacterium
- Burkholderia

C

- Campylobacter
- Candida
- Capnocytophaga
- Cardiobacterium
- Chryseobacterium
- Citrobacter
- Clostridium
- Comamonas
- Corynebacterium
- Cryptococcus

D

- Delftia

E

- Eggerthella
- Eikenella
- Elizabethkingia
- Enterobacter
- Enterococcus
- Erwinia
- Erysipelothrix
- Escherichia

F

- Finegoldia
- Fusobacterium

G

- Gamella
- Gardnerella
- Granulicatella

H

- Haemophilus
- Hafnia

K

- Kingella
- Klebsiella
- Kocuria
- Kodamaea

L

- Lactobacillus
- Leclercia
- Leptotrichia
- Leuconostoc
- Listeria

M

- Micrococcus
- Moraxella
- Morganella

N

- Neisseria
- Nocardia

O

- Oerskovia
- Oligella
- Orchobactrum

P

- Paenibacillus
- Pantoea
- Parabacteroides
- Paracoccus
- Parvimonas
- Pasteurella
- Peptoniphilus
- Peptostreptococcus
- Prevotella
- Propionibacterium
- Proteus
- Providencia
- Pseudomonas

R

- Raoultella
- Rhizobium
- Rhodotorula
- Roseomonas
- Rothia
- Ruminococcus

S

- Saccharomyces
- Salmonella
- Serratia
- Shigella
- Slackia
- Solobacterium
- Stenotrophomonas
- Streptococcus

T

- Trueperella

V

- Veillonella
- Vibrio

Y

- Yersinia

Text Book References for Microbank ®

The following text books reference the Microbank® Storage system as a recommended method:

Bailey & Scott's Diagnostic Microbiology, by P. Tille.
ISBN:9780323083300.

Laboratory Methods in Food Microbiology by W. F. Harrigan.
ISBN: 9780123260437

Fungal Plant Pathogens - Principles and Protocols Series by C. Lane, P. Beales, K. Hughes.
ISBN: 9781845936686

Probiotics in Food Safety and Human Health by I. Goktepe, V. K. Juneja, M. Ahmedna.
ISBN: 9781574445145

Cryopreservation and Freeze-Drying Protocols by J. G. Day, M. R. McLellan.
ISBN: 9780896032965

Manual of Techniques in Invertebrate Pathology by L. A. Lacey
ISBN: 9780123868992

Bergey's Manual of Systematic Bacteriology by W. Whitman, A. Parte, M. Goodfellow, P. Kämpfer, H-J. Busse, M. E. Trujillo, W. Ludwig, K.I. Suzuki.
ISBN: 9780387950433

Manual of Clinical Microbiology by J. Versalovic
ISBN: 9781555814632

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Microbank ®

If you have data that you'd like to share with Pro-Lab Diagnostics on the successful recovery of organisms using Microbank® please contact us.



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